



A-Form of Cytoplasmic Domain of nARIA (CRD-Neuregulin)
And uses Thereof

5 This application incorporates by reference the entire disclosure of U.S. Provisional Application No. 60/003,380, filed September 7, 1995 and U.S. patent application Serial No. 08/697,954, filed September 4, 1996.

10

A portion of the invention disclosed herein was made with Government support under NIH Grant No. NS29071 from the Department of Health and Human Services. Accordingly, the U.S. Government may have certain rights in this 15 invention.

Background of the Invention

Throughout this application, various publications are 20 referenced by author and date and by arabic numbers in superscript. Full citations for these publications may be found listed at the end of the specification immediately preceding the claims. The disclosures of these publications in their entireties are hereby 25 incorporated by reference into this application in order to more fully describe the state of the art as known to those skilled therein as of the date of the invention described and claimed herein.

30 The development and differentiation of embryonic neurons culminates in synapse formation. Neuronal development is an intricate process that involves a cascade of inductive interactions between a neuron and the pre- and postsynaptic partners of that neuron. These highly 35 regulated events are important for the establishment of reliable, yet plastic, synaptic formation and

transmission. Correct expression of an array of transmitter-gated channels by neurons is clearly essential to synaptic differentiation, and yet the developmental regulation of this process is poorly understood. In fact, despite overwhelming advances in probing the molecular and biophysical details of ion channels gated by gamma-amino butyric acid (GABA), glycine, glutamate and acetylcholine (ACh) (Betz, 1990; Deneris et al., 1991; McGehee et al., 1995; Role, 1992; Sargent, 1993) the corresponding embryonic versions of these receptors have evaded analysis. Characterization of the biophysical properties of ligand-gated channels in developing neurons and description of their evolution to the mature receptor profile is limited (Brussard et al., 1994; Moss and Role, 1993; Margiotta and Gurantz, 1989). Furthermore, little is known about the mechanism of these changes.

The study of embryonic ligand-gated channels and subsequent modifications of their functional profile during neural development is difficult. Receptor expression prior to synaptogenesis is at a low level. Synapse formation is not synchronous. In the few cases studied, the developmental changes in receptor function are vast (Berg et al., 1989; Engisch and Fischbach, 1992; Arenella et al, 1993; Deneris et al, 1991; McGehee and Role, 1995; Role, 1992; Sargent, 1993). In the establishment of mature synapses, profound alterations in the expression profile of neuronal ligand-gated channels occur. In addition to these changes in expression levels, changes in the cellular distribution, the subunit composition and the biophysical and pharmacological properties occur as well (Margiotta and Gurantz, 1989; Moss and Role, 1993; Moss et al., 1989; Devay et al, 1994; Arenella et al, 1993; Jacob, 1991; Mandelzys et al,

1994; Smith et al, 1983; Vernallis et al, 1993). The interactions between presynaptic and target neurons may play a large role in the extrinsic influences which are believed to modify receptor function throughout 5 development. The mechanism of receptor development remains unclear, however, presynaptic input, target cell regulation, synaptic activity or molecular signals independent of transmission may be involved.

10 **Diversity of neuronal nicotinic receptors**

One important feature of neuronal ligand-gated channels, nicotinic acetylcholine receptors (nAChRs) in particular, is the diversity of component subunits and the resultant diversity in channel subtypes (Boulter et al, 1986; 15 Conroy et al., 1992; Grynkiewicz et al., 1985; Lindstrom et al., 1990; Luetje and Patrick, 1991; McGehee and Role, 1995; Papke and Heinemann, 1991; Ramirez-Latorre et al., submitted; Role, 1992). Neuronal nAChRs were the first of the ligand-gated ion channels studied to display this 20 degree of structural and functional complexity. Although nAChRs comprise only two distinct subunit types, there are multiple homologous forms of each subunit encoding gene. There are 8 neuronal "α" subunit genes (α1-α8) and 3 neuronal "β" subunit genes (β2-β4) cloned to date 25 (Boulter et al., 1986; Heinemann et al., 1990; Nef et al., 1988; Seguela et al., 1993; Wada et al., 1989). With this array as a starting point, there could be more than 10^5 varieties of pentameric nAChR complexes (McGehee et al., 1995 and Role, 1992). Study of native nAChRs 30 indicates that the actual number of subunit combinations is less than theory would predict. Biochemical, immunochemical, and antisense deletion experiments to identify native compositions of nAChRs demonstrate that relatively few subunit combinations are likely to be 35 found in native nAChRs. For example, the nAChRs

expressed by autonomic and habenula neurons have been studied in detail (Brussard et al., 1994; Devay et al., 1994; Listerud et al., 1991; Clarke et al., 1986) and provide specific examples of the subunit composition of 5 each nAChR channel subtype expressed. In view of the documented evolution of these neuronal nAChR channels during embryonic development, and the array of molecular and biophysical tools available to study these channels in detail, an understanding of the developmental 10 regulation of nAChR subunit and channel subtype diversity may be close at hand. Numerous studies implicate the interaction during the formation of synaptic connections between the presynaptic and postsynaptic cells in the development of mature neuronal receptors (Arenella et 15 al., 1993; Boyd et al., 1988; Brussaard et al., 1994b; Brussaard et al., 1994; Devay (in preparation; Devay et al., 1994; Gardette, et al., 1991; Jacob 1991; Levey et al., 1994; Mandelzys et al., 1994; Moss et al., 1989).

20 **Regulation of neuronal phenotype during development:
contribution of target interactions.**

Neuronal differentiation is induced by the interaction of developing neurons with target cells. One example is that of the evolution of transmitter phenotype in a 25 special class of sympathetic neurons that evolve from an adrenergic to a cholinergic phenotype in the course of normal development. Although early on, these neurons synthesize, package and release catecholamines, the formation of synapses with the target sweat glands is 30 accompanied by a change in transmitter expression that ultimately produces a mature cholinergic phenotype. This change in transmitter expression requires both pre- and postsynaptic signals. Thus, catecholamine release from the embryonic neuron is required to induce the release of 35 a cell differentiation factor\leukemia inhibitory factor

(CDF/LIF)-like factor called sweat gland factor (SGF) from the presumptive sweat glands. SGF, released via activation of target adrenergic receptors, interacts, with specific receptors on the innervating neuron. SGF 5 induces the cellular machinery required for ACh synthesis and release in the presynaptic neuron. Thus, the attainment of a mature transmitter phenotype is regulated by both synaptic activity and target derived signals, offering an explanation for how the innervation of target 10 tissues regulates nAChR expression in CNS and PNS neurons.

This example is one of many implicating target-derived factors in the control of neuronal survival, 15 proliferation, differentiation, migration, and neurite outgrowth. Although there are many factors that could mediate target effects on neuronal differentiation, the expression patterns and biological activities of factors identified to date identify a few candidates for proposed 20 studies of nAChR regulation. (1) Ciliatory neurotropic factor (CNTF) mimics the effect of SGF in inducing a cholinergic phenotype. It has also been shown to promote differentiation of sympathetic precursor cells and likely participates in target-induced changes in nAChR 25 expression. CNTF is expressed in numerous sympathetic targets including smooth muscle and kidney. (2) Activin and related members of the Transforming Growth Factor β (TGF β) family, can also regulate the differentiation of the transmitter phenotype in autonomic neurons. These 30 factors are expressed in sympathetic targets such as smooth muscle, sweat glands etc. (3) CDF/LIF mimics SGF by inducing a cholinergic phenotype. This factor is secreted by smooth muscle and heart muscle. Although less is known about either the activity or the 35 distribution of these factors in the central nervous

system (CNS), it is likely that CNS and peripheral nervous system (PNS) neurons may be regulated by similar signaling molecules.

5 **Regulation of receptor synthesis and distribution in muscle: contribution of presynaptic signals**

Classical studies of Fischbach, Cohen, and McMahan of the nerve-muscle junction demonstrate that the incoming motor nerve is a potent regulator of muscle-nAChRs. Prior to 10 innervation, muscle expresses an embryonic form of nAChR which is diffusely distributed over the cell surface. The expression of the muscle-nAChR is eventually downregulated to a diffuse distribution. Elimination of muscle-nAChRs by innervation is accompanied by an 15 increase in local synthesis, insertion and formation of high-density clusters of muscle-nAChR at the synaptic site. At later stages of synaptic development, there are marked changes in the biological properties of muscle-nAChR channels due to alterations in subunit gene 20 expression. This produces "adult" type muscle-nAChR complexes of distinct subunit composition. Molecular signals that are believed to mediate these changes in muscle-nAChR distribution and synthesis have been identified and cloned, namely, agrin and AChR Inducing 25 Activity (ARIA). Recombinant agrin alters the distribution of pre-existent muscle-nAChRs with no effect on synthesis or insertion of new receptors. In contrast, recombinant ARIA induces muscle-nAChR subunit gene expression, increasing the rate of appearance of new 30 surface receptors from 3-5%/hr to 10-20%/hr.

It is possible that there are common regulatory mechanisms between nAChR and muscle-nAChR. It is believed that nAChRs on both CNS and PNS neurons evolve 35 from low density and diffuse distribution to clustered

and highly dense synaptic patches following innervation. Finally, like muscle-nAChRs, there are marked changes in the biophysical properties of nAChRs during development and presynaptic input may induce some of these changes, 5 e.g., channel conductance and opening frequency.

Despite the essential role of ligand-gated ion channels in synaptic transmission between neurons, little is known about changes in their expression, function, distribution 10 and subunit composition during neural development. Nicotine-induced enhancement of acquisition and consolidation of short term memories is believed to be mediated by presynaptic nAChR activation since this activation facilitates a broad array of CNS synapses. In 15 view of the impact that developmental changes in nAChRs have on neuronal excitability, synaptic efficacy and synaptic plasticity, studies of the regulatory controls of nAChR expression are essential. One avenue of study focuses upon the proteinaceous factors that appear to 20 modulate receptor gene expression. One factor previously identified and cloned is heregulin (See Vandlen and Holmes, U.S. Patent No. 5,367,060).

To date, 10 different proteins that result from 25 alternative splicing of the heregulin gene have been described. Among these are the growth factors neu differentiation factor (NDF) (Wen et al., 1992), glial growth factor (GGF) (Marchionni et al., 1993), ARIA (Falls et al, 1993; Fischbach et al., 1994), and the 30 heregulin isoforms (Holmes, 1992; Wen et al., 1994). The reported isoforms are principally membrane bound proteins which can be solubilized by proteolysis. The extracellular domains of these proteins consist of an N-terminal domain followed by an immunoglobulin-like (Ig-like) domain, a linker region, an EGF-like domain, and a 35

second linker region (Figure 5). These proteins are ligands for the epidermal growth factor (EGF) family of receptor tyrosine kinases. Binding of the ligand to the EGF receptor family members erbB3/HER3 or HER4 results in 5 activation of the tyrosine kinase activity of the receptor. Other family members can be activated by trans-phosphorylation via the activated members.

Other members of the heregulin/NDF/ARIA family have been 10 described in previous patent publications. PCT International Publication No. WO 94/08007, published April 14, 1994, entitled "Trophic factor having ion channel-inducing activity in neuronal cells" describes neurotrophic factors designated as ARIA, which are able 15 to induce the formation of ion channels. This publication also shows how ARIA is associated with both nervous tissue and skeletal muscle. ARIA has an Ig-like domain and an EGF-like domain in the extracellular region. U. S. Patent No. 5,367,060, issued November 22, 20 1994, entitled "Structure, production and use of heregulin", from U.S. Serial No. 847,743, filed March 6, 1992 by Richard L. Vandlen and William E. Holmes, discloses a polypeptide with a binding affinity for the p185^{HER2} receptor. Vandlen and Holmes also disclose 25 purification methods required to isolate heregulin and uses of the heregulins and antibodies specific to the heregulins as therapeutic agents. This polypeptide is related to but distinct from the ARIA protein.

Summary of the Invention

This invention provides an assay for diagnosing whether a subject has or is predisposed to developing a neoplastic disease which comprises: a) obtaining a biological sample from the subject; b) contacting the sample with an agent that detects the of an extracellular domain of nARIA (CRD-neuregulin) or an isoform thereof; c) measuring the amount of agent bound by the sample; d) comparing the amount of agent bound measured in step)c with the amount of agent bound by a standard normal sample, a higher amount bound by the sample from the subject being indicative of the subject being predisposed to developing a neoplastic disease. One embodiment of this invention is a method for maintaining synaptic connections between a neuron and a target cell comprising contacting the target cell with an nARIA polypeptide or a nucleic acid molecule encoding nARIA in an amount sufficient to induce the formation of a synaptic junction.

Brief Description of the Figures

Figure 1A-1C: Nucleotide sequence of chicken nARIA (SEQ ID NO: 1).

5 A nucleic acid sequence encoding a splice variant from the heregulin gene is shown. This sequence is a compilation of the sequences derived from the ExoIII deletion series on the pBluescript II KS (+) subclone of phage #3 from a chick E13 total brain cDNA library
10 screened with a rat pro-heregulin beta 1 probe generated by PCR amplification. The sequence was determined using the M13 reverse primer. The length is 3212 bases. The break in homology to the ARIA sequence occurs at a known splice site. The nucleotide sequences from base pair
15 number 1293 downstream to the poly-A tail of the nARIA clone are identical to ARIA. The sequences upstream from base pair number 1293 encode a unique splice variant (i.e. bp 1293-bp 3212), nARIA.

20 **Figure 2: Amino acid sequence of chicken nARIA (SEQ ID NO: 2).**

An amino acid sequence encoding a splice variant from the heregulin gene is shown. This sequence is a compilation of the sequences derived from the ExoIII deletion series
25 on the pBluescript II KS (+) subclone of phage #3 from a chick E13 total brain cDNA library screened with a rat pro-heregulin beta 1 probe generated by PCR amplification. The sequence was arrived at using the M13 reverse primer. The length is 1070 amino acids. The
30 asterisks denote unclear results these stop condons.

Figure 3: Nucleotide sequence of the unique portion of the human nARIA gene (SEQ ID NO: 3).

A nucleic acid sequence encoding the human nARIA (hnARIA)

a splice variant from the heregulin gene, nARIA is shown. The product was subcloned into pBluescript II KS (+) the 5' end of the transcript is at the M13 end of the multiple cloning site (MCS). The length is 1351 bases.
5 The unique portion of nARIA spans from base 93 to base 758.

Figure 4: Amino Acid sequence of the unique portion of the human nARIA protein (SEQ ID NO: 4).

10 An amino acid sequence encoding the unique portion of the human nARIA is shown. The product was subcloned into pBluescript II KS (+) the 5' end of the transcript is at the M13 end of the MCS. The length is 449 amino acids.

15 **Figure 5A-5B: Comparison of gene structure of nARIA with ARIA**

(A) Comparison of the gene structure of the chicken nARIA cDNA with ARIA human nARIA, heregulin, and NDF. (B) Comparison of the exon structure of splice variants of 20 the ARIA/NDF/Heregulin gene. The sequence we cloned (nARIA) has a unique N terminal domain devoid of Ig-like repeats common to other ARIA variants.

Figure 6: Stage E13 multiple tissue Northern blot.

25 Multiple tissue Northern blots were screened with probes specific for unique domains of nARIA and were compared with those probed with an ARIA specific probe. The ARIA probe indicates that this form is present in skeletal muscle (pectoral muscle) whereas expression of nARIA is 30 restricted to nervous tissue.

Figure 7: Developmental Northern analysis of ARIA and nARIA in the chick hindbrain and cerebellum

Northern blot analysis was performed on RNA samples from

chick embryonic stages E4 through E16. Oligonucleotides specific for either nARIA or ARIA were used as probes. (H-hindbrain; C-cerebellum.)

5 **Figure 8A-8B: RT-PCR analysis of the developmental expression pattern of nARIA in the chick brain compared with ARIA.**

(A) The developmental expression pattern of nARIA and ARIA in chick brain as detected by RT-PCR is shown. (B) 10 Relative quantification of ARIA and nARIA mRNA levels normalized with actin mRNA levels is shown.

15 **Figure 9A, 9B and 9C: nARIA and ARIA expression in the developing chick spinal cord as detected by PCR and Northern hybridization analysis.**

(A) Northern blot of ARIA (top) and nARIA (bottom). All probes and primers are directed against sequences specific to nARIA and/or ARIA. Northern analysis indicates the mRNA of nARIA is detectable by E3 and 20 robust by E4 whereas initiation of ARIA expression occurs later (e6-8) and is detectable at E6, but not robust until E8. (B) PCR detection of mRNA expression of ARIA and nARIA in developing chick spinal cord. Note specificity of primers tested on full length cDNAs (NA, 25 A) (-RT: no reverse transcriptase reaction; NA - nARIA positive control; A - ARIA positive control). (C) Comparison of nARIA and ARIA mRNA levels relative to actin.

30 **Figure 10A-10D: In situ hybridization of chick ED5 trunk cross-section of neural tissue with probes specific for nARIA and ARIA.**

The probe specific for nARIA contains the Cys-rich domain and the probe specific for ARIA contains the Ig domain.

Different patterns of expression are observed. A positive signal is observed in the presumptive preganglionic neurons with the nARIA probe but not with the ARIA probe. (A-D) ED5 trunk cross section. (A,C) 5 nARIA specific probe. (B,D) ARIA specific probe.

Figure 11A, 11B and 11C: nARIA induces tyrosine phosphorylation.

(A) Phosphorylation of MCF7 cell line demonstrating 10 activity of both recombinant ARIA and nARIA as ligands for tyrosine kinase-linked receptors. Dose (A) and time (C) dependence of ARIA (A) and nARIA (A,C) phosphorylations in MCF-7 cells. (B) Comparison of recombinant nARIA and ARIA tyrosine phosphorylation of 15 E9 lumbar sympathetic ganglia (LSG).

Figure 12: Anti Phosphotyrosine Western Blot of MDA-MB-453 cells treated with conditioned media.

Media conditioned by COS1 or HEK293 cells transiently 20 transfected with the nARIA clone (sense configuration) activates tyrosine kinase activity in the breast tumor cell lines MCF7 or MDA-MB-453 above the basal levels of tyrosine kinase activity (antisense configuration). The levels of phosphorylation of EGFR family members relative 25 to one another was different between ARIA and nARIA.

Figure 13A-13D: Electrophysiological assay of transmitter gated macroscopic currents.

(A,C) ACh-evoked current. (B,D) GABA-evoked response. 30 Treatment of primary cultures of sympathetic neurons from E11 chicks with recombinant nARIA for two days increases the magnitude of macroscopic currents activated by acetylcholine and appears to decrease the currents gated by GABA.

Figure 14A-14B: nARIA enhances expression of the nAChR subunit genes $\alpha 3$, $\alpha 5$, $\alpha 7$ and $\beta 4$ and increases the magnitude of $I_{P(ACh)}$.

(A) Assay of nAChR subunit gene expression in E9 neurons maintained *in vitro* for 3 days and then treated for 24 hrs with 10 μ l of recombinant nARIA (left) or ARIA (right) by quantitative RT-PCR. (B) Assay of $I_{P(ACh)}$ in E9 neurons maintained *in vitro* for 3 days and then treated for 48 hrs with 10 μ l of recombinant nARIA or ARIA or antisense construct of each (control). Macroscopic currents evoked by 500 μ M ACh. Peak current ($I_{P(ACh)}$) analyzed with non parametric tests appropriate for non-normally distributed values. Box-plots reveal the mid 50% of the data, whiskers delineate the 90% distribution. * indicates outliers. The nARIA induced increase and ARIA induced decrease in ($I_{P(ACh)}$) peak current are significant vs antisense controls at $p<0.01$, respectively.

Figure 15: Comparison of the affinity of nARIA vs ARIA for heparin sulfate proteoglycan.

MCF7 breast tumor cells were treated with conditioned media from either ARIA or nARIA transiently transfected COS1 cells. Some of the media was prebound with heparin attached to glass beads. Prior to treatment, the beads were pelleted by centrifugation to remove any heparin associated proteins. The supernatant was used to treat the MCF7 cells and tyrosine phosphorylation of the ARIA/nARIA receptor was analyzed.

30

Figure 16: The anti-nARIA antibody is specific for recombinant nARIA

Figure 17: Anti-nARIA antibodies reveal targeting of

nARIA to azon terminals at CNS and PNS synapses

Figure 18: nARIA induction of nAChR expression is more potent than the Ig-Containing isoforms.

5

Figure 19: Deletion of nARIA with isoform-specific antisense oligonucleotides reveals that nARIA is both necessary and sufficient for the regulation of nAChR expression by presynaptic input.

10

Figure 20: Disruption of CRD-NRG-1 gene products.

(A) Schematic diagram of β -form neuregulin-1 gene products (. β -forms differ from α -forms in the EGF-like domain). Splice variants of the NRG-1 gene and the various nomenclatures used in the literature are shown. 15 a,b,c refer to three variants of the cytoplasmic domain. Ext, extracellular, Tm, transmembrane, CYTO, cytoplasm.

(B) Schematic diagram of the wild-type CRD-containing allele, the targeting vector, and the mutated allele. The 20 barbell indicates the site of the nonsense mutation/Xba I site. Arrow and circled X schematically indicate that the nonsense mutation creates a translation stop codon. Note that the neo cassette is inserted in the antisense orientation into the intron 3' to the CRD-exon.

(C) Southern blot analysis of genomic DNA. Left. 25 ECO RI/Eco RV double digest, with probe A; Right, Xba I digest, with probe B.

(D) Southern Blot analysis of cDNA to examine neonatal neuregulin-1 transcripts. Top. Schematic representation 30 of CRD and Ig containing neuregulin-1 cDNAs. Arrowheads are per primers; solid bands are Ig and TM oligo probes used for Southern; hatched area is CRD-specific oligo probe. Note the presence of CRD-containing, Ig-containing, and EGF-TM-cytoplasmic containing transcripts

in CRD-NRG-1 ^(+/-) and CRD-NRG-1 ^(-/-) mice and that CRD transcripts are digested with Xba I only in CRD-NRG-1 ^(-/-) mice.

(E) a Newborn CRD-NRG-1 ^(-/-) mice lack spontaneous movement and are cyanotic. b,b' Whole-mount *in situ* hybridization with CRD-specific probe of E11 CRD-NRG-1 ^(+/-) & ^(-/-) mice. N5g, trigeminal; N10g, vagal ganglia; sc, spinal cord; drg, dorsal root ganglia. c,c' Paraffin-section of PO lung stained with H&E. (c) asterisks indicate expanded alveoli in CRD-NRG-T ^(+/-) mice, (c') carets point to collapsed airspaces in CRD-NRG-1 ^(-/-) mice.

Figure 21: Defective Neuromuscular Synaptogenesis in CRD-NRG-1 ^(-/-) mice.

(A) a,a' Immunofluorescence labeling (100X) of PO diaphragm with anti-neurofilament antibodies (red) or with b,b' anti-neurofilament antibodies (red) & α -bungarotoxin (α BgTx) (green). (a') arrows point to remnant neurofilament(+) staining in mutants, carets (b) and arrow(b') point to colocalization of neurofilament and α BgTx staining. c,c' Acetylcholinesterase staining (AChE) (400X) of PO diaphragm, (c) carets point to strong, aligned AChE staining in controls, (c¹) arrows point to weak, scattered AChE staining in mutant. d,d' H&E staining of fresh-frozen PO diaphragm muscle, (d) carets indicate eccentric, peripheral nuclei, (d') arrows point to central nuclei. (B) f,f' Immunofluorescence with anti-neurofilament antibodies (red); e,e',g,g' anti-neurofilament and anti-synaptophysin antibodies (red) & α BgTx (green); h,h' anti-neurofilament antibodies (green) & anti-S100 β antibody (red). (e,e') E18.5 intercostal muscle (400X), (e) asterisks point out overlap of neurofilament and α BgTx, (e') arrows indicate examples where staining does not overlap. f,f E14.5 diaphragm

(100X). (f) asterisks indicate normal branching pattern of phrenic nerve and termination of intradiaphragmatic branches in central region of muscle, (f) arrows highlight defasciculated axons with irregular branching patterns, repeated neurite crossings, and aberrant extension to the most lateral edges of the diaphragm. 5 (g,g') E14.5 diaphragm (400X). (g) asterisks indicate the organized endplate zone where nerve terminals overlie clusters of α BgTx-labeled AChRs. (g') arrows indicate 10 examples where synaptophysin & α BgTx do not overlap. Note disorganized endplate zone with nerve terminals approaching AChRs from many directions and some nerve terminals passing over α BgTx staining h.h' E14.5 diaphragm (400X). arrows in (h) indicate S100 β (+) cells 15 along peripheral nerves absent in (h').

Figure 22: Abnormal projections and subsequent degeneration of peripheral nerves in CRD-NRG-V $(-/-)$ mice.

(A) a,a' *In situ* hybridization with vesicular ACh 20 transporter (VAT)-specific probe of 10 μ m sections through cervical enlargement of E18.5 spinal cord (200X). b Number of motor neurons present in spinal cord levels C1-C6 in control and mutant embryos (n=3 for each age and genotype). c,c' Whole-mount immunostaining of E12.5 limb 25 with β -Tubulin III antibody (70X). (c) black and yellow asterisks show splayed nerve terminations of spinal nerves and cutaneous sensory projections in controls, respectively. (c') black and yellow arrows indicate bulbous, compact terminations of spinal nerves and 30 cutaneous sensory projections in CRD-NRG-V $(-/-)$ mice, respectively. **(B)** d,d' *In situ* hybridization with erbBS-specific probe of 10 μ m sections through cervical enlargement of E18.5 spinal cord (200X). d asterisks indicate sensory root lined with erbB3(+) cells, d' 35 arrows point to the sensory root that is devoid of

erbB3(+) cells. **e, e', f, f** Whole-mount immunostaining of E12.5 embryos with β -Tubulin III antibody (70X); Imb, limb. **(e, e')** asterisks and arrows indicate the cervical sensory cutaneous projections. **(f, f)** asterisks and arrows point to trunk lateral cutaneous sensory branches. **g, g'** Immunofluorescence with anti-VAT antibodies of E18.5 hindlimb muscle (green) (200X). Asterisks **(g)** & arrows **(g')** arrows point to nerve terminals. **h, h'** Whole-mount *in situ* hybridization of E12 embryos with erbBS-specific probe (70X). **(h)** asterisks point to erb3(+) staining along peripheral nerves emanating from caudal spinal cord levels, **(h')** arrows point to peripheral nerves which show decreased erbB3(+) staining. Note that embryo in **(h')** is older than embryo in **(h)**. **i, i'** Immunofluorescence with anti-neurofilament antibodies (green) & anti-S100 β antibody (red) (400X). **(i)** asterisks indicate colocalization of neurofilament & S100 β staining in nerve terminations of control, **(i')** yellow arrows point to colocalization of neurofilament and S100 β along preterminal branches; white arrows point to nerve terminations devoid of S100 β staining.

Figure 23: Hindbrain Deficits and Loss of Schwann cell precursors in CRD-NRG-1^(-/-) mice.

25 **(A)** **a-b** *In situ* hybridization of 10 μ m transverse serial sections of trigeminal ganglion of E14.5 embryos, **(a)** CRD-NRG-1 -specific probe (100X). **(b)** Pan-NRG-1 -specific probe (100X). **c-e** *In situ* hybridization of 10 μ m coronal serial sections of vibrissa follicles of E18.5 embryos, **(c)** CRD-specific probe (200X); **(d)** erbBS-specific probe (200X); **(e)** erbB4-specific probe (200X). **f, g** *In situ* hybridization of 10 μ m coronal sections of E18.5 embryos with CRD-NRG-1-specific probe, **(f)** trigeminal motor nucleus (200X) **(g)** facial motor nucleus (200X). **(B)** **h, h'**

Whole-mount immunostaining of E11 embryos (70X) with anti-p-tubulin III antibody, (h') Arrows indicate the onset of abnormal branching of cranial nerve projections N5n_{man}& N7n. N5g, trigeminal; N7g/N8g, vestibulo-cochlear; N9g, 5 petrosal; N10g, nodose ganglia; op, ophthalmic, max, maxillary, man, mandibular branches of N5n, trigeminal nerve; N7n, facial; N10n, vagus; N11n, spinal accessory; N12n, hypoglossal nerve; C1,C2 cervical dorsal root ganglion and cervical spinal cord projections. i,i' In 10 *situ* hybridization with erbBS-specific probe of trigeminal ganglion of E14.5 embryos, (i) asterisks indicate peripheral projections lined with erbB3(+) cells. T-bar indicates the width of nerve (100X). (i') arrows point to the peripheral nerve projection in 15 mutants (100X). (j,j') Whole-mount *in situ* hybridization with erbBS- specific probe of E12 embryos (70X). (j) black and green arrows point to erbBS-positive cells along N11n and its convergence with N10n and along peripheral nerve projections of cervical dorsal root 20 ganglia, respectively. White arrows point to erbB3(+)positive cells in developing muscle, (j') black and green arrows point to the dramatic loss of erbB3(+) staining along N11n, the coalescence of N10n, and along the projections of cervical dorsal root ganglia, 25 respectively. White arrowhead points to the remaining erbBS staining in developing muscle in CRD-NRG-^(-/-) mice. k,k',l,l' *In situ* hybridization of E18.5 coronal sections through hindbrain with a vesicular ACh transporter (VAT)- specific probe. Note the reduction in size of facial 30 motor nucleus (50X) (k vs k') and of trigeminal motor nucleus (200X) (I vs I') and abnormal cell migration (arrows, k') in CRD-NRG-1 ^(-/-) mice. Aq, aqueduct of IVth ventricle, m, midline, LOT, lateral dorsal tegmental nucleus; PBr, parabrachial nucleus; N6n, abducens nucleus;

-20-

N7n, facial nucleus. Dotted line in \mathbf{k}, \mathbf{k}' indicates the midline, m.

Detailed Description of the Invention

This invention provides an isolated nucleic acid molecule encoding nARIA. This nucleic acid molecule may encode 5 human nARIA (hnARIA), wherein the nucleic acid comprises the sequence shown from base 93 to base 758 of Figure 3 (SEQ ID NO: 3). This invention also provides for an isolated nucleic acid molecule encoding nARIA, wherein the nucleic acid molecule encodes chicken nARIA (cnARIA) 10 which comprises the sequence shown from base 608 to base 1234 of Figure 1 (SEQ ID NO: 1). The nucleic acid molecule may be DNA, cDNA or RNA. The isolated nucleic acid molecule encoding nARIA includes nucleic acids encoding functionally equivalent variants or mutants of 15 nARIA including nucleic acid molecules which, due to the degeneracy of the genetic code, code for nARIA polypeptide such as the polypeptide shown in Figures 2 (SEQ ID NO: 2) and 4 (SEQ ID NO: 4).

20 The isolated nucleic acid molecule encoding nARIA includes nucleic acids encoding biologically active variants of nARIA. This includes nucleic acid molecules which are capable of specifically hybridizing with an nARIA sequence. Biologically active variants may include 25 nucleic acids variants which have at least 75% amino acid sequence identity with an nARIA sequence, more preferably at least 80%, even more preferably at least 90% and most preferably at least 95%. Identity or homology with respect to an nARIA sequence is defined herein as the 30 percentage of amino acid residues in the candidate sequence that are identical with nARIA residues in Figure 2 (SEQ ID NO: 2) and 4 (SEQ ID NO: 4), after aligning the sequences and introducing gaps, if necessary, to achieve the maximum percent homology, and not considering any 35 conservative substitutions to be identical residues.

None of N-terminal, C-terminal or internal extensions, deletions, or insertions into nARIA sequence shall be construed as affecting homology. The isolated nucleic acid molecule encoding nARIA also includes any splice 5 variants having nARIA biological activity as defined hereinafter.

As used herein, the purified nARIA polypeptide includes biologically active nARIA polypeptides which include each 10 expressed or processed nARIA sequence, fragments thereof having a unique consecutive sequence of at least 5, 10, 15, 20, 25, 30 or 40 amino acids residue as shown in the underlined regions in Figure 2 (SEQ ID. NO: 2) and 4 (SEQ ID. NO: 4). Biologically active amino acids residue has 15 been inserted N- or C- terminal to, or within the nARIA sequence. Amino acid sequence variants include nARIA wherein an amino acid residue has been replaced by another residue, nARIA polypeptides including those containing predetermined mutations by, e.g. site-directed 20 or PCR mutagenesis. nARIA includes nARIA from such species as rabbit, rat, porcine, non-human primate, Drosophila, equine, murine, opine, human and chicken and alleles or other naturally occurring variants of the foregoing; derivatives of nARIA wherein it has been 25 covalently modified by substitution, chemical, enzymatic or other appropriate means with a moiety such as an enzyme or radioisotopes. nARIA may be labeled with a detectable moiety including a fluorescent label, a biotin, a digozigenin, a radioactive atom, a paramagnetic 30 ion, and a chemiluminescent label. This invention also provides for glycosylation variants of nARIA (as in the insertion of a glycosylation site or deletion of any glycosylation site by deletion insertion or substitution of an appropriate residue); and soluble forms of nARIA, 35 such as nARIA which lacks a functional transmembrane

domain.

As used herein, the purified nARIA polypeptide includes amino acid variants of nARIA which are prepared by 5 introducing appropriate nucleotide changes into nARIA nucleic acid or by *in vitro* synthesis of the desired nARIA polypeptide. Such variants include, for example, deletions from, or insertions or substitutions of, residus within the amino acid sequence shown for human 10 nARIA sequence. Any combination of deletions, insertion, and substitution of, residues within the amino acid sequence shown for human nARIA sequence. Any combination of deletions, insertion, and substitution can be made to arrive at the final construct possesses the desired 15 characteristics. The amino acid changes also may alter post-translational modifications of nARIA, such as changes in the glycosylation sites, altering the membrane anchoring characteristics, altering the location of nARIA by inserting, deleting or otherwise affecting the 20 transmembrane sequence of native nARIA or modifying its susceptibility to proteolytic cleavage.

This invention also provides for fusion proteins which contain nARIA polypeptide linked to an unrelated protein 25 domain(s). The fusion proteins may be created by the insertion of amino- and/or carboxyl-terminal fusions ranging in length from one residue to polypeptides containing a hundred or more residues, as well as intrasequence insertions of single or multiple amino acid 30 residues. Intrasequence insertions (i.e. insertions within the nARIA sequence) may range generally from about 1 to 10 residues, or preferably 1 to 5, and most preferably 1 to 3. Examples of terminal insertions include nARIA with an N-terminal methionyl residue (an 35 artifact of the direct expression of nARIA in bacterial

recombinant cell culture), and fusion of a heterologous N-terminal signal sequence to the N-terminus of nARIA to facilitate the secretion of mature nARIA from recombinant host cells. Such signal sequences generally will be 5 obtained from, and thus be homologous to, the intended host cell species. Suitable sequences include STII or 1 pp for *E. coli*, alpha factor for yeast, and viral signals such as herpes gD for mammalian cells.

10 Other insertional variants of nARIA may include the fusion of the N- or C-terminus to an immunogenic polypeptide, e.g., bacterial polypeptides such as beta-lactamase or an enzyme encoded by the *E. coli* *trp* locus, or yeast protein, bovine serum albumin, or chemotactic 15 polypeptides.

As used herein, the purified nARIA polypeptide includes amino acid substitution variants. These variants have at least one amino acid residue in the nARIA molecule 20 removed and a different residue insert in its place. The sites of greatest interest for substitutional mutagenesis include a site(s) identified as an active site(s) of nARIA, and sites where the amino acids found in nARIA ligands from various species are substantially different 25 in terms of side-chain bulk, charge, and/or hydrophobicity.

The amino terminus region of the cytoplasmic region of the nARIA may be fused to the carboxy terminus of 30 heterologous transmembrane domains and receptors, to form a fusion polypeptide useful for intracellular signalling of a ligand binding to the heterologous receptor.

Other sites of interest are those in which particular 35 residues of nARIA-like ligands obtained from various

species are identical. These positions may be important for the biological activity of nARIA. These sites, especially those falling within a sequence of at least three other identically conserved sites, are substituted 5 in a relatively conservative manner. Such conservative substitutions may be known as "preferred substitutions" and may include: valine substituted for alanine; lysine for arginine; glutamine for asparagine; glutamate for aspartate; serine for cysteine; asparagine for glutamine, 10 aspartate for glutamate; proline for glycine; arginine for histidine; leucine for isoleucine; arginine for lysine, leucine for methionine, leucine for phenylalanine; glycine for proline, threonine for serine; serine for threonine; tyrosine for tryptophan; 15 phenylalanine for tyrosine and leucine for valine.

Substantial modifications in function or immunological identity of nARIA are accomplished by selecting substitutions that differ significantly in their effect 20 on maintaining (a) the structure of the polypeptide backbone in the area of the substitution, for example as a sheet or helical conformation, (b) the charge or hydrophobicity of the molecule at the target site, or (c) the bulk of the side-chain. Naturally occurring residues 25 are divided into groups based on common side chain properties: hydrophobic; neutral hydrophilic; acidic; basic; residues that influence chain orientation and aromatic. Non-conservative substitutions will entail exchanging a member of one of these classes for another. 30 Such substituted residues may be introduced into regions of nARIA that are homologous with other receptor ligands, or, more preferably, into the non-homologous regions of the molecule.

35 This invention provides for the creation of a

combinatorial library of potential nARIA homologs which can be generated from a degenerate oligonucleotide sequence. Chemical synthesis of a degenerate gene sequence can be carried out in an automatic DNA 5 synthesizer, and the synthetic genes can then be ligated into an appropriate vector for expression. The synthesis of degenerate oligonucleotides is well known in the art (see Sambrook, et al., 1989; U.S. Patents Nos. 5,223,409; 5,198,346 and 5,096,815. The purpose of making such a 10 library is to provide in one mixture, all of the sequences encoding the desired set of potential nARIA sequences. This mixture could then be used for selection of particular affinities, binding properties and separate functionalities.

15

This invention also provides for a replicable vector which contains nARIA sequence and a host cell containing this vector. This expression vector may be a prokaryotic expression vector, a eukaryotic expression vector, a 20 mammalian expression vector, a yeast expression vector, a baculovirus expression vector or an insect expression vector. Examples of these vectors include PKK233-2, pEUK-C1, pREP4, pBlueBacHisA, pYES2, PSE280 or pEBVHis. Methods for the utilization of these replicable vectors 25 may be found in Sambrook, et al., 1989 or in Kriegler 1990. The host cell may be a eukaryotic cell, a somatic cell, a germ cell, a neuronal cell, a myocyte, a mammary carcinoma cell, a lung cell, a prokaryotic cell, a virus packaging cell, or a stem cell.

30

This invention also provides for a purified polypeptide or an isolated polypeptide encoding nARIA. This polypeptide may encode human nARIA protein which includes the sequence shown from amino acid 31 to amino acid 252 35 of Figure 4 (SEQ ID NO: 4). This polypeptide may also

encode chicken nARIA protein which includes the sequence shown from amino acid 203 to amino acid 421 of Figure 2 (SEQ ID NO: 2). This invention provides for the genomic sequence of human nARIA and chicken nARIA. This 5 invention provides for functionally equivalent variants or mutants of nARIA including polypeptides which contain replacement amino acids which do not affect the functionality of the polypeptide. These variants may be prepared using *in vitro* mutagenesis techniques, 10 polymerase chain reaction mutagenesis, or site-directed mutagenesis. The invention also provides for nARIA derivatives *in vitro* which are immobilized on a support for purposes of diagnoses, purification of nARIA binding factors or affinity purification of nARIA antibodies.

15

According to the present invention, nARIA or neuronal Acetylcholine Receptor Inducing Activity possesses certain biological activities. As used herein, nARIA has the ability to increase both nAChR currents and nAChR 20 subunit gene expression in LSG neurons. The sequence of nARIA is unique throughout the N-terminal portion of the sequence, lacking the Ig-like domain typically upstream of the EGF-like domain in ARIA. nARIA retains the juxtamembrane EGF-like domain, shown to be sufficient for 25 receptor binding and activation of receptor tyrosine kinase activity. Expression of nARIA by Northern blot analysis begins at E4 and is maximal at E8, while expression of ARIA begins at E6 and peaks at E8 in the chick embryo. In cultured E11 chick sympathetic neurons 30 treated with cultured media from COS cells transiently transfected with either ARIA or nARIA, an assay of both acetylcholine (ACh) gated currents and subunit mRNA levels was performed. These assays demonstrated differential regulation of nAChRs by nARIA vs ARIA. 35 Specifically, nARIA significantly increased the maximal

responses to 500 mM ACh whereas ARIA significantly decreased the maximal responses compared to cultures treated with recombinant protein from the antisense construct. Measurement of nAChR subunit mRNA levels in 5 E9 sympathetic neurons treated with nARIA or ARIA with quantitative RT-PCR revealed different profiles of subunit gene regulation. A 24 hour treatment with nARIA mimicked the effects of innervation, up-regulating α_3 , α_5 , α_7 and β_4 levels, whereas ARIA downregulated β_4 and 10 α_3 . nARIA, therefore, may participate in the increase in nAChR subunit transcription induced by innervation of embryonic sympathetic neurons *in vivo* during sympathetic neuronal development.

15 nARIA, as discussed herein, not only has a unique n-terminal region but also displays biological activity distinct from ARIA. Tales I, II and III in the Experimental Details section herein describe these distinctions. nARIA, unlike ARIA, is specifically 20 expressed only in nervous tissue, whereas ARIA can be expressed in other tissues. nARIA expression is higher in the spinal cord and cerebellum than in the forebrain and optic tectum. nARIA expression is first detected at stage E4 in the spinal cord, and expression is first 25 detected in the cerebellum at stage E8. In all E11 sympathetic neurons, nARIA specifically has an effect on ligand gated channels: in response to acetylcholine, nARIA specifically increases the number of functional acetylcholine receptors as indicated by an increase in 30 response to maximal concentrations of acetylcholine. nARIA has little effect on the number of GABA activated channels as indicated by the response to maximal concentrations of GABA. In contrast, at E11, ARIA has little effect on acetylcholine evoked responses and may 35 upregulate GABA evoked responses. The effects of nARIA

and ARIA on the acetylcholine evoked responses indicates that nARIA has been quantitated at E9 and is about 15 times more potent than ARIA.

5 nARIA increases the transcription of the α_3 subunit of nAChR in sympathetic neurons. nARIA also increases α_5 , α_3 , α_7 and β_4 subunit gene expression of nAChR.

This invention provides for a method of inducing the
10 expression of a specific nicotinic acetylcholine receptor subunit isoform. AChRs at mature mammalian neuromuscular junctions are pentameric protein complexes composed of four subunits in the ratio of $\alpha_2\beta\gamma\delta$ (Mishina et al 1986). Most, if not all, of embryonic AChRs contain a different
15 subunit, termed "tau" in place of the epsilon subunit. When mixtures of α, β, δ and tau subunit mRNAs are injected into *Xenopus* oocytes, the expressed channels have the properties of embryonic receptors. It is likely that this change in subunit composition is due to a change in
20 gene expression and accounts for the switch in properties of ACh-activated channels from slow channels to fast channels. This invention provides for the application of nARIA alone or in combination with another agent to neural cells to induce the expression of subunit
25 isoforms.

This invention further provides for nARIA antagonists which are capable of reducing the biological activity of nARIA. This antagonist may be proteinaceous such as an
30 antibody specific for nARIA as described herein, a nucleic acid such as an antisense molecule to the nARIA mRNA as described herein, an enzymatic activity such as a ribozyme directed to nARIA mRNA as described herein or a protease specific for the nARIA polypeptide. The
35 antagonist may also be an agent which is capable of

binding the nARIA receptor with higher affinity than nARIA, thus competing away the effects of nARIA binding.

The subject invention also provides for nARIA agonist(s) which would be capable of enhancing the biological activity of nARIA. Such agonists may include other neurotrophic factors such as ciliary neurotrophic factor (see U.S. Patent No. 4,997,929); nerve growth factor (see U.S. Patent No. 5,169,762); neurotrophic factor 4/5 (see PCT International Publication No. WO 92/05254); brain-derived neurotrophic factor (see U.S. Patent No. 5,180,820); glial-derived neurotrophic factor (see PCT International Publication No. WO 93/06116) or any other neurotrophic factor (see European application EP 0 386 752 A1). The disclosures of these publications in their entireties are hereby incorporated by reference into this application in order to more fully describe the state of the art as known to those skilled therein as of the date of the invention described and claimed herein.

20

This invention also provides for an antibody immunoreactive with an epitope comprising a unique sequence shown in either Figure 2 (SEQ ID NO: 2) from amino acid 203 to amino acid 421 or in Figure 4 (SEQ ID NO: 4) from amino acid 31 to amino acid 252.

A further embodiment of the invention is a monoclonal antibody which is specific for nARIA. In contrast to conventional antibody (polyclonal) preparations which typically include different antibodies directed against different determinants (epitopes), each monoclonal antibody is directed against a single determinant on the antigen. Monoclonal antibodies are useful to improve the selectivity and specificity of diagnostic and analytical assay methods using antigen-antibody binding. Also, they

may be used to remove nARIA from the serum. A second advantage of monoclonal antibodies is that they can be synthesized by hybridoma cells in culture, uncontaminated by other immunoglobins. Monoclonal antibodies may be 5 prepared from supernatants of cultured hybridoma cells or from ascites induced by intraperitoneal inoculation of hybridoma cells into mice. The hybridoma technique described originally by Köhler and Milstein, 1976, has been widely applied to produce hybrid cell lines that 10 secrete high levels of monoclonal antibodies against many specific antigens.

Another embodiment of this invention is a ribozyme which is capable of cleaving nARIA mRNA. See Cech, et al., 15 U.S. Patent No. 4,987,071; Altman et al., U.S. Patent No. 5, 168,053; Haseloff et al, U.S. Patent No. 5,254,678 published European application No. Hampel et al., EP 360,257.

20 This invention provides for a nucleic acid comprising a unique nARIA sequence in a 3' to 5' orientation, antisense to at least a portion of a gene encoding naturally occurring nARIA. This antisense nucleic acid molecule may be labeled with a detectable moiety selected 25 from the group consisting of a fluorescent label, a biotin, a digoxigenin, a radioactive atom, a paramagnetic ion, and a chemiluminescent label. See Inoue et al. U.S. Patent Nos. 5,208,149 and 5,190,931 and Schewmaker, U.S. Patent No. 5,107,065.

30 Labeling of a circular oligonucleotide (such as a replicable vector as described herein) can be done by incorporating nucleotides linked to a "reporter molecule" into the subject circular oligonucleotides. A "reporter 35 molecule", as defined herein, is a molecule or atom

which, by its chemical nature, provides an identifiable signal allowing detection of the circular oligonucleotide. Detection can be either qualitative or quantitative. The present invention contemplates using 5 any commonly used reporter molecule including radionucleotides, enzymes, biotins, psoralens, fluorophores, chelated heavy metals, and luciferin. The most commonly used reporter molecules are either enzymes, 10 fluorophores, or radionucleotides linked to the nucleotides which are used in circular oligonucleotide synthesis. Commonly used enzymes include horseradish peroxidase, alkaline phosphatase, glucose oxidase and α -galactosidase, among others. The substrates to be used with the specific enzymes are generally chosen because a 15 detectably colored product is formed by the enzyme acting upon the substrate. For example, p-nitrophenyl phosphate is suitable for use with alkaline phosphatase conjugates; for horseradish peroxidase, 1,2-phenylenediamine, 5-aminosalicylic acid or toluidine are commonly used. The 20 probes so generated have utility in the detection of a specific nARIA DNA or RNA target in, for example, Southern analysis, Northern analysis, *in situ* hybridization to tissue sections or chromosomal squashes and other analytical and diagnostic procedures. The 25 methods of using such hybridization probes are well known and some examples of such methodology are provided by Sambrook et al, 1989. This invention also provides a method of amplifying a nucleic acid sample comprising priming a nucleic acid polymerase chain reaction with 30 nucleic acid (DNA or RNA) encoding (or complementary to) an nARIA.

Another embodiment of this invention is the normal expression or overexpression of nARIA *ex vivo* in human 35 neuronal cells, stem cells or undifferentiated nerve

cells and muscle cells. These cells may be utilized for gene therapy in patients (See Anderson et al U.S. Patent No. 5,399,346).

5 This invention further provides for a transgenic nonhuman mammal whose germ or somatic cells contain a nucleic acid molecule which encodes nARIA polypeptide or biologically active variants thereof, introduced into the mammal, or an ancestor thereof, at an embryonic stage. This
10 invention provides for a transgenic nonhuman mammal whose cells may be transfected with a suitable vector with an appropriate sequence designed to reduce expression levels of nARIA polypeptide below the expression levels of that of a native mammal. The transgenic nonhuman mammal may
15 be transfected with a suitable vector which contains an appropriate piece of genomic clone designed for homologous recombination. Alternatively, the transgenic nonhuman mammal may be transfected with a suitable vector which encodes an appropriate ribozyme or antisense
20 molecule. See for example, Leder and Stewart, U.S. Patent No. 4,736,866 for methods for the production of a transgenic mouse.

Biologically functional variants of nARIA are nucleic
25 acid molecules that, due to the degeneracy of the genetic code, code on expression for nARIA polypeptide. The foregoing variant DNA sequences may be translated into variant nARIA polypeptides which display the biological activity of an nARIA polypeptide. These variant nucleic acid molecules may also be expressed in this transgenic mammal. Active variants should hybridize to the wild-type nARIA nucleic acid sequence under highly stringent or moderately stringent conditions (Sambrook et al, 1989).

One embodiment of this invention is a method for inducing the formation of a synaptic junction between a neuron and a target cell, which includes treating the target cell with nARIA polypeptide or nARIA nucleic acid molecule 5 encoding nARIA or a biologically active variant thereof, in an amount sufficient to induce the formation of a synaptic junction. A "sufficient amount" as used herein refers to that amount which provides a therapeutic effect for a given condition and administration regimen. The 10 target cell may be a somatic cell such as a myocyte, a neuronal cell, a glandular cell or any postsynaptic cell. This method provides for the induction of the formation of a synaptic junction in an individual having a neurological disorder involving abnormal synaptic 15 connections. Isolated nARIA may be used as a growth factor for *in vitro* cell culture or *in vivo* to promote the growth of cells.

nARIA, nARIA agonists or nARIA antagonists may be used to 20 treat any disease where levels of nARIA metabolism are changed and therefore ion channel levels or activities are not normal as in some neurological disorders. Such disorders would include any disease with an abnormal production of nARIA. Neurological disorders that affect 25 the central nervous system, memory or cognitive functions may also be treated with nARIA. Such disorders may be the result of the normal aging process or the result of damage to the nervous system by trauma, surgery, ischemia, infection or metabolic disease. Such disorders 30 may also include Alzheimer's Disease, Turret's Syndrome, and Parkinson's Disease. These diseases have been shown to respond to nicotine treatment.

The neurological disorder may be a neuromuscular 35 disorder. Examples of neuromuscular disorders which may

be treatable with nARIA include Alzheimer's disease, myasthenia gravis, Huntington's disease, Pick's disease, Parkinson's disease, and Turret's Syndrome. Also included are neurogenic and myopathic diseases including 5 chronic atrophies such as amyotrophic lateral sclerosis, Guillain-Barre syndrome, progressive bulbar palsies, spinal muscular atrophies and chronic peripheral neuropathy. Autonomic disorders of the peripheral nervous system may also be included in this treatment 10 which include disorders that affect the innervation of muscle or endocrine tissue such as tachycardia, atrial cardiac arrhythmias and hypertension. These disorders are thought to be associated with an abnormally low level 15 of muscarinic AChRs in the striated muscle.

15

This invention provides for a method of altering neuro-receptor expression. In this method, nARIA is administered to a subject which may result in a change in 20 the expression of neuro-receptors. The method of administration of nARIA is described more fully hereinafter.

This invention provides for the production of functional mammalian nARIA protein in a prokaryotic expression 25 system, a mammalian expression system, a baculovirus expression system, an insect expression system or a yeast expression system. This production may provide for the post-translational modifications which exist in the naturally occurring nARIA protein. For protocols 30 describing bacterial expression of mammalian proteins, see Sambrook et al, 1989.

Another embodiment of this invention is a method for inducing neuronal regeneration which comprises contacting 35 a target cell with a composition of nARIA and a

pharmaceutically acceptable carrier to induce the formation of a synaptic junction between a neuron and a target cell. The target cell may be a neuronal cell, an endocrine cell, a muscle cell or any cell capable of 5 forming a neuro-muscular junction. nARIA may be used to facilitate incorporation of implants into nervous tissue or to promote nerve regeneration following damage by trauma, infarction, infection or postoperatively.

10 This invention provides for a combination therapy of nARIA with another neurotropic factor or cytokine or growth factor or with other agents known for use in the treatment of malignancies. Such factors may include transforming growth factor beta (TGF- β), ciliary 15 neurotropic factor (CNTF), brain derived neurotropic factor (BDNF), NT-4, NT-5, NT-4/5, nerve growth factor (NGF), activins, agrin, cell differentiation factor (CDF), glial growth factor (GGF), and neu differentiation factor (NDF), ARIA, and heregulins. nARIA may be 20 administered in combination with agrin for effects on the neuromuscular junction. For therapy directed toward the autonomic/enteric nervous system, TGF- β and nARIA is the preferred combination. For therapy directed to the central nervous system, nARIA and CDF is the preferred 25 combination.

When administered parenterally, proteins are often cleared rapidly from the circulation and may therefore elicit relatively short-lived pharmacological activity. 30 Consequently, frequent injections of relatively large doses of bioactive proteins may be required to sustain therapeutic efficacy. Proteins modified by the covalent attachment of water-soluble polymers such as polyethylene glycol, copolymers of polyethylene glycol and 35 polypropylene glycol, carboxymethyl cellulose, dextran,

polyvinyl alcohol, polyvinylpyrrolidone or polyproline are known to exhibit substantially longer half-lives in blood following intravenous injection than do the corresponding unmodified proteins (Abuchowski et al., 5 1981; Newmark et al., 1982; and Katre et al., 1987). Such modifications may also increase the protein's solubility in aqueous solution, eliminate aggregation, enhance the physical and chemical stability of the protein, and greatly reduce the immunogenicity and 10 antigenicity of the protein. As a result, the desired *in vivo* biological activity may be achieved by the administration of such polymer-protein adducts less frequently or in lower doses than with the unmodified protein. nARIA compositions may be administered 15 parenterally by injection or directly into the cerebral spinal fluid by continuous infusion from an implanted pump. nARIA may also be administered with one or more agents capable of promoting penetration of nARIA across the blood-brain barrier.

20 Attachment of polyethylene glycol (PEG) to proteins is particularly useful because PEG has very low toxicity in mammals (Carpenter et al., 1971). For example, a PEG adduct of adenosine deaminase was approved in the United 25 States for use in humans for the treatment of severe combined immunodeficiency syndrome. A second advantage afforded by the conjugation of PEG is that of effectively reducing the immunogenicity and antigenicity of heterologous proteins. For example, a PEG adduct of a 30 human protein might be useful for the treatment of disease in other mammalian species without the risk of triggering a severe immune response. nARIA or cells that produce nARIA may be delivered in a microencapsulation devise so as to reduce or prevent an host immune response 35 against the nARIA producing cells. nARIA may also be

delivered microencapsulated in a membrane, such as a liposome.

Polymers such as PEG may be conveniently attached to one
5 or more reactive amino acid residues in a protein such as
the alpha-amino group of the aminoterminal amino acid,
the epsilon amino groups of lysine side chains, the sulfhydryl
groups of cysteine side chains, the carboxyl
groups of aspartyl and glutamyl side chains, the alpha-
10 carboxyl group of the carboxy-terminal amino acid,
tyrosine side chains, or to activated derivatives of
glycosyl chains attached to certain asparagine, serine or
threonine residues.

15 Numerous activated forms of PEG suitable for direct
reaction with proteins have been described. Useful PEG
reagents for reaction with protein amino groups include
active esters of carboxylic acid or carbonate
derivatives, particularly those in which the leaving
20 groups are N-hydroxysuccinimide, p-nitrophenol, imidazole
or 1-hydroxy-2-nitrobenzene-4-sulfonate. PEG derivatives
containing maleimido or haloacetyl groups are useful
reagents for the modification of protein free sulfhydryl
groups. Likewise, PEG reagents containing amino
25 hydrazine or hydrazide groups are useful for reaction
with aldehydes generated by periodate oxidation of
carbohydrate groups in proteins.

Another embodiment of this invention is a method for
30 determining a prognosis of or diagnosing a neoplastic
condition in a subject. In this method, one may obtain a
biological sample from the subject, and contact the
sample with a reagent capable of binding to an element in
the sample, the element being an nARIA nucleic acid
35 molecule or polypeptide encoding nARIA, under conditions

such that the reagent binds only if the element is present in the sample. One may then detect the presence of the reagent bound to the element and thereby determine the prognosis of the neoplastic condition of the subject.

5 The reagent may be an oligonucleotide capable of hybridizing with a nucleic acid encoding nARIA polypeptide under standard stringency hybridization conditions. The reagent in this method may be an antibody specific for nARIA polypeptide. The element in
10 the biological sample may be a nucleic acid molecule encoding nARIA or a polypeptide encoding nARIA protein. The biological sample may be cerebrospinal fluid, blood, plasma, ascites fluid, tissue, urine, sputum, amniotic fluid, saliva, lung lavage, or cell extracts. This
15 method may be performed with the reagent is affixed to a solid support. The neoplastic condition may be a mammary neoplasm or a small cell carcinoma of the lung.

A further embodiment of this invention is a method for
20 the treatment of a neoplastic condition of a subject. In this method a pharmaceutically acceptable form of nARIA in a sufficient amount over a sufficient time period is administered to a subject to induce differentiation of neoplastic cells and thus treat the neoplastic condition.

25 The composition may be a form of nARIA such as nARIA polypeptide or nARIA nucleic acid, combined with a pharmaceutically acceptable carrier. The carrier may be made up of suitable diluents, preservatives, solubilizers, emulsifiers, or adjuvants and may be in an
30 aerosol, intravenous, oral or topical form.

Also provided by the invention are pharmaceutical compositions comprising therapeutically effective amounts of polypeptide products of the invention together with
35 suitable diluents, preservatives, solubilizers,

emulsifiers, adjuvants and/or carriers. A "therapeutically effective amount" as used herein refers to that amount which provides a therapeutic effect for a given condition and administration regimen. Such 5 compositions are liquids or lyophilized or otherwise dried formulations and include diluents of various buffer content (e.g., Tris-HCl., acetate, phosphate), pH and ionic strength, additives such as albumin or gelatin to prevent absorption to surfaces, detergents (e.g., Tween 10 20, Tween 80, Pluronic F68, bile acid salts), solubilizing agents (e.g., glycerol, polyethylene glycerol), anti-oxidants (e.g., ascorbic acid, sodium 15 metabisulfite), preservatives (e.g., Thimerosal, benzyl alcohol, parabens), bulking substances or tonicity modifiers (e.g., lactose, mannitol), covalent attachment 20 of polymers such as polyethylene glycol to the protein, complexation with metal ions, or incorporation of the material into or onto particulate preparations of polymeric compounds such as polylactic acid, polyglycolic acid, hydrogels, etc, or onto liposomes, microemulsions, 25 micelles, unilamellar or multilamellar vesicles, erythrocyte ghosts, or spheroplasts. Such compositions will influence the physical state, solubility, stability, rate of *in vivo* release, and rate of *in vivo* clearance of nARIA. The choice of compositions will depend on the 30 physical and chemical properties of the protein having nARIA activity. For example, a product derived from a membrane-bound form of nARIA may require a formulation containing detergent. Controlled or sustained release 35 compositions include formulation in lipophilic depots (e.g., fatty acids, waxes, oils). Also comprehended by the invention are particulate compositions coated with polymers (e.g., poloxamers or poloxamines) and nARIA coupled to antibodies directed against tissue-specific receptors, ligands or antigens or coupled to ligands of

tissue-specific receptors. Other embodiments of the compositions of the invention incorporate particulate forms protective coatings, protease inhibitors or permeation enhancers for various routes of administration, including parenteral, pulmonary, nasal, oral, injection or infusion by intravenous, intraperitoneal, intracerebral, intramuscular, intraocular, intraarterial or intralesional. nARIA may be part of a pharmaceutical composition with agrin and an acceptable carrier to recapitulate both the induction of expression of AChR and the clustering of the AChR's on the membrane surface.

Polypeptides of the invention may be "labeled" by association with a detectable marker substance (e.g., radiolabeled or biotinylated) to provide reagents useful in detection and quantification of nARIA or its receptor bearing cells in solid tissue and fluid samples such as blood or urine.

Another embodiment of this invention is a method for determining whether a compound is capable of modulating the binding of an nARIA polypeptide to its receptor. In this method, the compound may be incubated under suitable conditions with an appropriate nARIA polypeptide-affinity derivative or receptor-affinity derivative under appropriate conditions such that an affinity complex may form. Then, one may measure the amount of affinity complex formed so as to determine whether the compound is capable of modulating the binding of the nARIA polypeptide to its receptor. The affinity complex may be an nARIA receptor bound to an affinity derivative or an nARIA polypeptide bound to a derivative. The measurement in this method may comprise binding of an antibody specific for nARIA to the affinity complex to measure the

amount of affinity complex formed. The affinity derivative may be sepharose, cellulose, plastic, glass, latex, glass beads, a nylon membrane, a cellulose acetate membrane, an epoxy-activated synthetic copolymer 5 membrane, a nitrocellulose membrane or a streptavidin-coated plastic.

The present invention provides an assay for diagnosing whether a subject has or is predisposed to developing a 10 neoplastic disease which comprises: a) obtaining a biological sample from the subject; b) contacting the sample with an agent that detects the presence of an extracellular domain of nARIA (CRD-neuregulin) or an isoform thereof; c) measuring the amount of agent bound 15 by the sample; d) comparing the amount of agent bound measured in step c) with the amount of agent bound by a standard normal sample, a higher amount bound by the sample from the subject being indicative of the subject having or being predisposed to developing a neoplastic 20 disease.

The present invention also provides assay for determining whether a subject has a neurodegenerative disease which comprises: a) obtaining a biological sample from the 25 subject; b) contacting the sample with an agent that detects the presence of an extracellular domain of nARIA (CRD-neuregulin) or an agent which detects the presence of soluble neuregulin receptor; c) measuring the amount of agent bound by the sample; d) comparing the amount of 30 bound agent measured in step c) with the amount of agent bound by a standard normal sample, a higher amount or a lower amount bound by the sample from the subject being indicative of the subject having a neurodegenerative disease.

In one embodiment, the agent is an antibody or a fragment thereof. In another embodiment, the sample is cerebrospinal fluid (CSF), blood, plasma, sputum, amniotic fluid, ascites fluid, breast aspirate, saliva, 5 urine, lung lavage, or cell lysate or extract derived from a biopsy. In a further embodiment, the agent is an antibody which binds to an epitope formed by the amino acid sequence shown in Figure 2 (SEQ ID NO: 2) or Figure 4 (SEQ ID NO: 4). In another embodiment, the agent is an 10 antibody which binds to an epitope of the cytoplasmic domain of nARIA. In another embodiment, the agent is an antibody which specifically binds to the amino acid sequence NQDPIAV (SEQ ID NO: 5). In a further embodiment, the neoplastic disease is breast cancer, 15 prostate cancer, a brain cancer or ovarian cancer. In another embodiment, the neurodegenerative disease is Alzheimer's Disease, Parkinson's Disease, Turrets Syndrome, Amyotrophic lateral Sclerosis, Pick's Disease, Myasthenia Gravis, or Senility.

20 The present invention also provides a method for maintaining or sustaining a synaptic connection between a neuron and a target cell comprising contacting the target cell with an nARIA polypeptide or a nucleic acid 25 molecule encoding nARIA or biologically active variant thereof, in an amount sufficient to maintain the synaptic junction.

In one embodiment, the target cell is a somatic cell 30 including a myocyte, a neuronal cell, a glandular cell or any postsynaptic cell. In another embodiment, maintenance of the synaptic junction is accomplished in an individual having a neurological disorder involving abnormal synaptic connections. In another embodiment, 35 the neurological disorder is an neuromuscular disorder or

a neurodegenerative disease. In a further embodiment, the nARIA polypeptide is NQDPIAV (Seq ID NO: 5) or the A-form of the cytoplasmic domain of nARIA. In a further embodiment, the neurological disorder is Alzheimer's 5 Disease, Parkinson's Disease, Turrets Syndrome, Amyotrophic Lateral Sclerosis, Pick's Disease, Myasthenia Gravis, or Senility.

The present invention also provides a method for inducing 10 neuronal regeneration which comprises contacting a target cell with a composition of nARIA and pharmaceutically acceptable carrier to induce the formation of a synaptic junction between a neuron and a target cell. In one embodiment, the target cell is a neuronal cell or a 15 muscle cell.

The present invention also provides a method for the treatment of a neoplastic condition of a subject which comprises administering to the subject a pharmaceutically 20 acceptable form of nARIA in a sufficient amount over a sufficient time period to induce differentiation of neoplastic cells and thus treat the neoplastic condition.

25 The present invention also provides a method for determining whether a compound is capable of modulating the binding of an nARIA polypeptide to its receptor, which comprises: (a) incubating the compound under suitable conditions with an appropriate nARIA 30 polypeptide-affinity derivative or receptor-affinity derivative under appropriate conditions such that an affinity complex may form; (b) measuring the amount of affinity complex formed so as to determine whether the compound is capable of modulating the binding of the 35 nARIA polypeptide to its receptor.

In one embodiment, the affinity complex comprises an nARIA receptor bound to an affinity derivative. In another embodiment, the affinity complex comprises an nARIA polypeptide bound to an affinity derivative. In 5 another embodiment, the measuring comprises binding of an antibody specific for nARIA to the affinity complex to measure the amount of affinity complex formed. In another embodiment, the affinity derivative comprises sepharose, cellulose, plastic, glass, glass beads, or a 10 streptavidin-coated plastic.

The present invention also provides an assay for detecting neoplastic disease in a subject which comprises: a) obtaining a biological sample from the 15 subject; b) contacting the sample with an agent that specifically binds to an expression product of a neuregulin gene or a neuregulin receptor; c) measuring the amount of agent bound by the sample; d) comparing the amount of agent bound measured in step c) with the amount 20 of agent bound by a standard normal sample, a higher amount bound by the sample from the subject being indicative of the presence of neoplastic disease in the subject.

25 In one embodiment, the neuregulin receptor is erbB2, erbB3, or erbB4. In another embodiment, the agent specifically binds to an amino acid sequence of neuregulin which directs translocation to the nucleus. In another embodiment, an expression product of an 30 neuregulin gene comprises a neuregulin protein, an extracellular domain of a neuregulin protein, a polypeptide encoded by the amino acid sequence shown in Figure 2 (SEQ ID NO: 2) or 4 (SEQ ID NO: 4). In another embodiment, the agent is detectably labeled.

The present invention provides an assay for determining the amount of her 2 receptor in a bodily fluid from a subject and an assay for other heregulin receptors which comprises a) obtaining a bodily fluid from a subject or 5 a sample from a subject (i.e. a brain punch biopsy, blood, serum, plasma, urine, etc.); b) contacting the sample with cells which are transfected with a reporter construct (the construct being CRD-neuregulin with a cytoplasmic tail which has a detectable marker attached to 10 it); c) measuring the detectable label in the nucleus of the cells and thereby determining the amount of her 2 receptor in the bodily fluid from the sample.

The neuregulin in the bodily fluid binds to a receptor on 15 the surface of the transfected cell and there will be cleavage of the external domain and the cleavage of the internal cytoplasmic domain. The cytoplasmic domain will then be translocated into the nucleus and the detectable 20 label will be available for detection in the nucleus. The detectable label may be green fluorescent protein or VP16.

Another assay which is provided for in the present invention is an assay which comprises obtaining a brain 25 punch from a subject and performing immunohistochemistry on frozen sections or preserved sections of the tissue obtained using a detectably labeled antibody specific for the cytoplasmic domain of neuregulin and determining the percentage of nuclei that are showing label in the 30 tissue sample.

The nuclear targeting domain of neuregulin may be found in Yang et al and begins at amino acid number 230 (KTKKQRKK).

This invention is illustrated in the Experimental Detail section which follows. These sections are set forth to aid in an understanding of the invention but are not intended to, and should not be construed to, limit in any 5 way the invention as set forth in the claims which follow thereafter.

Experimental Details

Example 1 - Isolation and Sequence Analysis of nARIA

5 Novel members of the ARIA/NDF/hereregulin family that would
be expressed in neurons projecting to cholinoreceptive
neural targets were identified. Cloning efforts yielded
29 positives, including several which encoded variants
with an entirely novel N-terminal sequence, distinguished
10 by the absence of the usual Ig-like domains of the
hereregulin/NDF family. The predominance of the novel, Ig-
less clones over the ARIA like clones was striking (11 vs
5 of 29 positives). We named the novel splice variant
15 "nARIA" for **n**euronal **AChR** **I**nducing **A**ctivity. All clones
were isolated by two different and separate approaches as
described below.

A different library was used in each screening protocol
carried out to isolate and clone the nARIA gene. A chick
20 E13 total brain cDNA library was screened with a rat DNA
probe generated by PCR amplification. For the PCR
amplification, degenerate primers corresponding to
nucleotide sequences 523-542 (upper primer) and 1080-1100
25 (lower primer) of the published rat NDF sequence (Wen et
al., 1992) were used to amplify a DNA fragment from a
template of adult rat spinal cord cDNA. The upper and
lower primers were within the immunoglobulin and
transmembrane domains respectively. The amplified
fragment was subcloned into the pCRII[®] vector (Invitrogen)
30 and was sequenced, revealing an open reading frame. The
predicted peptide encoded by this fragment contained the
immunoglobulin to transmembrane domains of the hereregulin
β1 isoform and is distinct from the published NDF
sequence, which is an α-isoform of hereregulin. Screening
35 of the cDNA library by random primed labeling of the

amplified fragment resulted in three independent clones, of which only one contained a complete open reading frame. This open reading frame is 2055 nucleotides (Figure 1 (SEQ ID NO: 1), nucleotides 608-2662) and 5 encodes the nARIA transcript. The nucleotide sequences from base pair 1293 downstream to the poly-A tail of the nARIA clone are identical to ARIA a related cloned chicken gene (Falls et al., 1993).

10 On the protein level, identical sequences encode the portion of the molecule spanning from the EGF-like domain to the C-terminus in ARIA and nARIA. The break in homology occurs at a known splice site and the sequences upstream to the splice junction are unique to nARIA.
15 Analysis of the predicted protein sequence did not produce a motif corresponding to an immunoglobulin domain. Instead, there was a cysteine rich region identified (8 cysteines in 34 amino acids). See Figure 2 (SEQ ID NO: 2).

20

Example 2 - Spatial and Temporal Expression of nARIA in chick development

A chick E5-E11 spinal cord cDNA library which we prepared 25 was also screened. The probe for screening was generated by RT-PCR amplification from E8 chick spinal cord total RNA using primers corresponding to nucleotide sequences 264-281 (upper primer) and 1294-1313 (lower primer) of the published ARIA sequence (Falls, et al., 1993). The 30 amplified PCR fragment was subcloned into a PGEM3Z vector and sequenced to confirm its identity. Screening of the chick E5-11 spinal cord primary cDNA library by random primed labeling using the PCR fragment as the template resulted in 26 clones. Of these, 11 contained the novel 35 cysteine rich domain (nARIA like clones), 6 clones

contained the immunoglobulin-like domain (ARIA like splice forms), while the remaining clones had unidentifiable sequences. The nARIA clones included 8 that were identical to the form obtained by the earlier 5 screening and 3 additional isoforms that differed in the N-terminus, the EGF-like domain, the juxtamembrane linker, and/or the C-terminal region. The extracellular domains of these clones were fully sequenced and the intracellular portions were partially sequenced. 10 Restriction mapping and Southern blotting were also used to confirm the relatedness of these molecules. A human cerebellar cDNA library was screened in the manner described above. This procedure resulted in the isolation of human nARIA (Figures 3 (SEQ ID NO: 3) & 4 15 (SEQ ID NO: 4)).

Multiple tissue Northern blots were screened with probes specific for unique domains of nARIA and were compared with those probed with an ARIA specific probe (Figure 6). 20 In particular, the ARIA probe detected ARIA in skeletal muscle (pectoral muscle) whereas expression of nARIA was found to be restricted to nervous tissue. The expression of nARIA represents a higher percentage of the total message in the cerebellum and spinal cord than in the 25 forebrain or optic tectum. ARIA message is represented at a higher level in the forebrain and optic tectum than in the cerebellum and spinal cord (Figure 7). The developmental expression patterns of nARIA and ARIA in spinal cord as detected by RT-PCR and Northern blot 30 hybridization are different (Figure 8A-8B). The mRNA of nARIA is detectable by E3 and robust by E4 whereas initiation of ARIA and expression occurs later (E6-8) (Figure 9A-C).

35 *In situ* hybridization studies with probes specific for

nARIA and ARIA (containing the cysteine rich domain or Ig-like domain respectively) also demonstrated different patterns of expression (Figure 10A-D). In particular, a positive signal is obtained in the presumptive 5 preganglionic neurons with the nARIA probe but not with the ARIA probe. Therefore, the pattern of expression of ARIA and nARIA are different.

Example 3 - Functional Analysis of Biological Activity -
10 nARIA Activates Tyrosine Kinase Linked Receptor(s).

Initial experiments to characterize the functional properties of the nARIA protein focused upon the ability to activate protein tyrosine kinases. ARIA has been 15 proposed to act, as other members of the NDF/hereregulin family, through an interaction with specific tyrosine kinase-linked receptors (Falls, et al., 1993). This first step in transduction is assayed as tyrosine phosphorylation of a high molecular weight band, thought 20 to represent phosphorylation of the receptor subunit(s). We examined the pattern of tyrosine phosphorylated proteins in extracts of lumbar sympathetic ganglia (LSG) neurons as well as several other cell lines with an anti-TYR-P antibody (4G10; UBI) (Ausubel et al., 1994; Falls, 25 et al., 1993). Both recombinant ARIA and NARIA (from transiently transfected COS cells) induced time and dose dependent phosphorylation of 170-185 kD bands in the MCF7 and MDA-MB-453 cell lines (human carcinomas that overexpress erb-B2 receptor). ARIA appeared more potent 30 than nARIA and somewhat less robust in phosphorylation of a 185 kD band in LSG. Differential effects of nARIA on glial cells and neurons differ from all other hereregulin isoforms including ARIA, examined to date. nARIA's unique N-terminal sequence influences the binding of the 35 isoform to the protein tyrosine kinase receptors thereby

conferring distinct specificities.

Media conditioned by COS1 or HEK293 cells transiently transfected with the nARIA clone (sense configuration) 5 activated tyrosine kinase activity in the breast tumor cell lines MCF7 or MDA-MB-453 above the basal levels of tyrosine kinase activity as determined by using the antisense configuration of nARIA. See Figure 11A-C. Furthermore, the levels of phosphorylation of the EGFR 10 family members relative to one another was different between ARIA and nARIA. ARIA treatment resulted in a higher level of erb3/HER3 phosphorylation than nARIA. Treatment of acutely dispersed sympathetic neurons from E9 chicks with nARIA conditioned media resulted in 15 increased tyrosine phosphorylation of an approximately 180 kD protein (Figure 12).

Another assay to more clearly delineate between the biological activities of nARIA and ARIA involved the 20 comparison of their effects on the expression of ligand-gated channels in primary neurons. These studies assayed the number of functional surface receptors for two transmitters (ACh and GABA) using an electrophysiological assay of transmitter gated 25 macroscopic currents. The rationale for these experiments is based upon the pattern of expression of nARIA and ARIA, and on our previous studies of receptor regulation by spinal cord neurons (Role, 1988; Gardette et al, 1991). Treatment of primary cultures of 30 sympathetic neurons from E11 chicks with recombinant nARIA for two days increased the magnitude of macroscopic currents activated by acetylcholine and appeared to decrease the currents gated by GABA (Figure 13A-D). In contrast, treatment with recombinant ARIA under the same 35 conditions decreased the currents gated by acetylcholine

and appeared to enhance GABA-evoked currents. The differential effects of nARIA vs ARIA on ACh gated currents, an index of the number of functional channels on the cell surface, was also reflected in the assays of 5 the levels of expression of ACh receptor subunit encoding mRNA's.

Notably, a 24 hour treatment with recombinant nARIA increased the level of $\alpha 3$ subunit mRNA; in contrast, the 10 level of $\alpha 3$ subunit mRNA was either slightly decreased or not altered by ARIA. The differential effect of nARIA and ARIA on transcription was not limited to the acetylcholine receptor subunits. Application of the differential display technique to primary cultures of 15 sympathetic neurons treated for 24 hours with nARIA or ARIA suggested that the two growth factors differentially activate or suppress transcription of several distinct cDNAs.

20 The data presented herein suggests that the novel splice variant of the heregulin gene, nARIA may play a role in synaptic development that is unique from that of ARIA or other immunoglobulin-domain-containing splice variants. nARIA may potentially be used therapeutically or 25 diagnostically. Alterations in the level of the production of nARIA may be indicative of a traumatic insult to the nervous system. Since the receptor for this factor is a known oncogene, changes in growth factor levels may be prognostic to some neoplastic conditions. 30 Recombinant nARIA may be useful in cancer treatment regimens or for use in neuronal regeneration as described more fully herein. Other isoforms have been demonstrated to induce differentiation of breast tumor cell lines, promote survival of glial cells and increase the 35 mitogenesis of some cell lines.

It has been demonstrated that the biological activity of nARIA is different than that of ARIA and is summarized in Table I. (The bold serves to highlight the differences in biological activity between ARIA and nARIA.)

Table I. Biological Activity and Effects of ARIA vs nARIA

| Biological Activities | ARIA | nARIA |
|------------------------------------|--|---|
| 5 Expression | <p>Seen in chick nervous tissues and skeletal muscle</p> <p>Expression is higher in the forebrain and optic tectum than in the spinal cord and cerebellum</p> <p>Expression starts at E6 in the spinal cord</p> <p>Expression starts at E8 in the cerebellum</p> | <p>Seen only in nervous tissue at E13</p> <p>Expression is higher in the spinal cord and cerebellum than in the forebrain and optic tectum</p> <p>Expression starts at E4 in the spinal cord</p> <p>Expression starts at E8 in the cerebellum</p> |
| Activation of p185 phosphorylation | <p>Seen in L6 rat myocytes</p> <p>Seen in PC12 cells (Fischbach, et al)</p> <p>Seen in sympathetic neurons</p> <p>Seen in ciliary neurons (Fischbach, et al)</p> <p>Seen in breast tumor cell lines MCF7 and MDA-MB-453</p> <p>Seen in rat O2A cells (Fischbach)</p> | <p>Seen in L6 rat myocytes</p> <p>Not seen in PC12 cells</p> <p>Seen in sympathetic neurons</p> <p>Not determined</p> <p>Seen in breast tumor cell lines MCF7 and MDA-MB-453</p> <p>Not determined</p> |

| | | | |
|----|---|--|--|
| | Effects on ligand gated channels | Increases response to acetylcholine in E9 and decreases response in E11 sympathetic neurons Increases response to GABA in E9 and increases response in E11 sympathetic neurons. Increases response to ACh in muscle cells (Fischbach et al.) | Increases response to acetylcholine in E9 and increases response in E11 sympathetic neurons Increases GABA responses in E9 and decreases responses in E11 sympathetic neurons Not determined |
| 5 | Effects on transcription of nAChR subunits in rat medial habenula | No effect | Not determined |
| 10 | Effects on transcription of nAChR subunits in sympathetics | No effect on $\alpha 3$ Increases $\alpha 5$ and $\alpha 7$ | Increases $\alpha 3$ Increases $\alpha 5$ and $\alpha 7$ |
| 15 | Effects on sodium channel in muscle | Increases (Fischbach et al.) | Not determined |
| 20 | Effects on PC12 differentiation | Induces very short neurites in 10% of the cells (Fischbach et al.) Reduces rate of replication by half (Fischbach et al.) | Not determined Not determined |
| | Effects on glial cells | Increases the number of oligodendrocytes that develop from O2A precursors (Fischbach et al.) | Not determined |

| | | |
|---|-----------------------------------|----------------|
| Effects on nAChR subunits in PC12 cells | "Like control" (Fischbach et al.) | Not determined |
|---|-----------------------------------|----------------|

5

Macroscopic, Single channel and synaptic current data acquisition and analysis

Single channel data acquisition and analysis was 10 performed with an Axon Instruments system using Axobasic and PCLAMP 6.0 software. Additional programs were specifically designed for resolution of multiple channel classes of similar size and kinetics. Conductance, 15 kinetics, NPO and mean I analyses were performed as previously described (Listerud et al., 1991; Moss and Role, 1993; Moss et al., 1989; Simmons et al., 1988). Continuously recorded and evoked synaptic and macroscopic currents were stored on videotape and analyzed off line in software written in Axobasic.

20

An 80486 DX2-66Mhz computer equipped with the Axobasic system was essential to all studies. The acquisition program sampled all events that conformed to the amplitude and rise-time criteria, both set up by the 25 user. Each captured trace included 20 msec of pre-event baseline data. The system sampled events accurately up to 20 Hz--entirely adequate for capturing the relatively low frequency events in the experiments described herein. The analysis software provides amplitude, frequency, 30 rise- and decay-time constant information for each current recorded.

Subsequent generation of histograms, cumulative plots fitting, and statistical analyses were performed with 35 Microsoft Excel 3.0, SigmaPlot 4.1 (Jandel Scientific) and Systat. Synaptic current frequency information was

divided into bins for plotting and statistical comparisons. Statistical analyses of differences between control and treatment groups were evaluated by a two-tailed test (Snedecor and Cochran, 1989). Synaptic 5 current amplitude data were compared by plotting cumulative histograms. These plots were also utilized as estimated cumulative probability distributions for the determination of statistically significant differences between treatment groups using the Komolgorov-Simirnov 10 test (Press et al., 1986).

Table II provides a comparison of synaptogenesis vs. recombinant nARIA or ARIA treatment in regulating nAChR gene expression. (Bold serves to highlight the 15 differences.)

Table II. Comparison of Synaptogenesis vs. Recombinant nARIA or ARIA treatment in regulating nAChR gene expression.

| AChR subunit | α2 | α3 | α4 | α5 | α7 | β2 | β4 |
|--|-----------|---------------------------------|------------|-------------|-------------|-----------|---------------------------|
| Development <i>in vivo</i> | ND | 170% | ! to ND | 500% | 1000% | +/- | 600% |
| Presynaptic input <i>in vitro</i> | -- | 240% | -- | 315% | 150% | -- | 195% |
| Target contact <i>in vitro</i> Heart----- Kidney----- | -- | !; 60% of control 148% | -- | 189% | 261% | ? | no ^a |
| recombinant N-ARIA | -- | 224% | -- | 248% | 372% | ? | [0- 400%] |
| recombinant ARIA | -- | !; 90% of control | -- | 265% | 215% | ? | no ^a |

Table III provides a comparison of the regulation of ACh-gated currents induced by input and target vs nARIA and ARIA.

5 **Table III. Comparison of the Regulation of ACh-Gated Currents Induced by Input & Target vs. nARIA and ARIA.**

| | <u>Presynaptic input in vitro</u> | | <u>Target Contact in vitro:</u> | |
|----|-----------------------------------|-----------|---------------------------------|------------------|
| 10 | SMN | ↑1300% | Heart | ↓ 30% of control |
| | VMN | ↑750% | | |
| 15 | <u>Input Cond. Media</u> | | <u>Target Cond. Media</u> | |
| | SMN | ↑420% | Heart | ↓ 50% of control |
| 20 | VMN | ↑430% | Kidney | ↓ 225% |
| | recombinant nARIA | ↑200-400% | recombinant ARIA | ↓60% of control |

25 **Experimental Methods**

Cell Culture

LSG co-cultures with presynaptic input and target.

Dissociated embryonic sympathetic neurons from ED10 and ED17 were prepared and maintained *in vitro* as described 30 in Role 1988 with modifications as noted herein. Under these conditions the neurons were devoid of nonneuronal cells and were both adrenergic and cholinceptive. Innervation of sympathetic neurons by preganglionic microexplants was done according to previously described 35 techniques (Gardette et al. 1991; Hasselmo and Bower, 1993). Assay of target effects on nAChR expression required coculture of atrial micro-explants (ED12) with LSG neurons *in vitro*. Changes in expression of subunit mRNAs were assayed after 3-4 days of coculture by

quantitative RT-PCR (see below and Habecker and Landis, 1994).

Patch Clamp Recording

5 Recording of macroscopic and synaptic currents employed the whole-cell tight seal recording configuration of the patch clamp technique (Hammil et al., 1981). This techniques provided low noise recordings that allowed for resolution of elemental synaptic currents. Fabrication
10 of patch electrodes, pipette and bath solutions were all as previously described (Moss and Role, 1993; Moss et al., 1989). Currents were recorded with an AXOPATCH 200A patch clamp amplifier and stored on videotape with a PCM digitizer (Instrutech VR-10B) for subsequent analysis off
15 line. $I_{P(AChR)}$ is peak current.

Drug Application

Drugs and agonists were applied by microperfusion to small groups of cells via a large barreled delivery tube
20 with continuous macroperfusion at 1 ml/min. This approach optimized speed of application (<30 msec), speed of removal and the ease of changing test solutions applied by the same device. A stable perfusion set-up was an essential component of the each recording set up.

25

Molecular Techniques

Identification of subunit gene expression by PCR

The profile of subunit gene expression was analyzed by
30 PCR amplification of cDNA using nAChR subunit specific primers according to our previously published techniques (Listerud et al., 1991). Briefly, total RNA was extracted by homogenization of tissue in 4M guanidine thiocyanate buffer followed by centrifugation through a
35 5.7M CsCl cushion. The isolated RNA was DNase treated

and reverse transcribed using oligo-dT primers. AMV-RT(reverse transcriptase) was used to amplify a fragment encoding the most variable portion of the nAChR subunits, the intracellular loop. The identity of the amplified 5 products was verified by restriction mapping and/or Southern blotting (Ausubel et al., 1994).

Quantitative RT-PCR

Cell contents were collected by aspiration into DEPC 10 containing solution and cDNA was synthesized by addition of random hexamer primers and Superscript[®] reverse transcriptase enzyme (BRL). The cDNA served as template for amplification by primers specific for the various nAChR subunits. The internal standard construct included 15 sequences complimentary to all upstream and downstream primers used with an interposed multicloning site (MCS) linker. Thus, the efficiency of primer annealing to standard and to the cDNA template was equivalent. In order to detect the product of the reaction, trace 20 amounts of isotope labeled nucleotides were added to the reaction mixture. After 23 rounds of amplification, an aliquot of the reaction mixture was removed and further amplified with fresh Taq polymerase and reaction mix. This step was repeated after another 23 cycles. The 25 amplified fragment was separated from the unincorporated nucleotides by electrophoresis and the product was quantitated. The assay provided subunit specific quantification of α 2, α 3, α 4, α 5, α 7, α 8, β 2 and β 4 in 30 individual samples which detected as little as 2 fg of each subunit.

COS cell transfection

Cells were tansfected according to established techniques (Falls et al., 1993). Briefly, pcDNA1-amp containing the 35 nARIA or ARIA cDNA in sense or antisense orientation was

introduced into COS cells using lipofectamine_® (Gibco-BRL) per manufacturer's instructions. Twenty-four (24) hrs after transfection, the cells were washed and incubated in serum-free OPTI-MEM_® (Gibco-BRL). After 48 hrs, the 5 media was collected, centrifuged to remove debris and then concentrated 22-X using a Centriprep_® 10 concentrator (Amicon). Aliquots were stored at -20°C until use.

Tyrosine-phosphorylation assay

10 Cells were treated with L-15 media plus concentrated conditioned media from either sense, antisense, or non-transfected COS cells. After the desired incubation time, the cells were washed and lysed in 1% NP-40 buffer. The lysate was centrifuged and the protein in the 15 supernatant was quantified with a Bradford analysis procedure (BioRad). Protein samples were electrophoresed on a 4% SDS-PAGE gel and electroblotted onto a PBDF membrane (S&S Inc). Then the membrane was probed for phosphotyrosine with a monoclonal antibody 4G10 (UBI), 20 detected with a peroxidase-conjugated anti-mouse IgG2_b antiserum (Boehringer-Mannheim) and visualized using the luminescent ECL_® reagents (Amersham).

In situ hybridization

25 Cell-specific expression of AChR subunit mRNAs were assayed using *in situ* hybridization in tissue sections of spinal preganglionic nuclei with ³⁵S-labeled complementary RNA probes as described (Devay et al., 1994; Ausubel et al, 1994). cDNAs encoding the non-conserved regions 30 between transmembrane-spanning regions, TM3 and TM4, for each subunit have been subcloned into pGEM-3Z plasmids. Antisense riboprobes were transcribed in the presence of ³⁵S-UTP using the Promega transcription kit. Hybridization was assayed by autoradiography, and Nissl 35 staining allowed visualization of the cell bodies. ³⁵S-

labeled RNA probes provided good signal resolution with low background due to the high specific activity (cpm/probe molecule) and also due to the relatively low energy of ^{35}S emission. Determination of hybridization 5 involved comparison between parallel assays with antisense cRNA, sense RNA and RNase pretreatment of the tissue. Non-isotopic labeling protocol utilizing Digoxigenin-11-UTP in place of the ^{35}S -UTP was also used. The protocols were very similar (Bertrand et al., 1991).

10

Antisense oligonucleotide design and experimental protocols

Antisense oligonucleotides for AChR subunits $\alpha 2$, $\alpha 3$, $\alpha 4$, $\alpha 5$ and $\alpha 7$ were targeted to a 15 base sequence spanning 15 the initiation site of each subunit mRNA. The region upstream and including the ATG was divergent among the subunit sequences and in no case included <4 base mismatch with all chick cDNA sequences registered in GCG. Control oligos included missense sequences of identical 20 composition of oligos mutated at 3 of the 15 bases (same GCG ratio). The uptake, metabolism, hybridization and block of subunit expression by oligonucleotides were studied in some detail to determine optimal conditions for specific block. Briefly, neurons were pretreated 25 with an irreversible nAChR ligand (bromoacetylcholine bromide; BAC) and then incubated for 6-48 hrs with 10 μM oligo in heat inactivated medium (Gardette et al., 1991; Listerud et al., 1991). The d-oligos were taken up and intact 15-mer within the cells was maximal within 6 hrs 30 and still detected up to 48 hrs. $\alpha 2$, $\alpha 3$, $\alpha 4$, $\alpha 5$ and $\alpha 7$ have all been studied with this technique to determine their contribution to medial habenula nucleus (MHN) and LSG somatic nAChRs with reliable functional block by 24-48 hours. Antisense oligonucleotides were also designed 35 to inhibit the expression of nARIA and ARIA. These

oligos were directed against the translation start site or sequence within the N-terminal domain since the sequences are maximally divergent in this region. To optimize the antisense mediated block, and minimize the 5 confounding contribution of pre-existent nARIA and ARIA, antisense was introduced just prior to the initial surge in expression of these factors during development (\approx ED4 for nARIA; \approx ED8 for ARIA). Treatment of preganglionic tissue of these ages with the antisense oligos 10 effectively knocked out the expression of these factors and allowed us to ascertain the extent to which each of these factors contributed to developmental changes in nAChR expression.

15 A question remains as to whether ARIA or nARIA is required for regulation of nAChR channels by presynaptic input. This idea is tested by the selective block of nARIA or ARIA synthesis by antisense-mediated deletion. To optimize the antisense treatment, the region 20 containing the presynaptic neurons is removed prior to the initial surge in nARIA and ARIA expression (E4 and E8, cord; E4 septal region). In this manner we may succeed in blocking the major increase in expression, thereby obviating effects of pre-existent nARIA or ARIA. 25 Presynaptic microexplants are treated for 24-48 hrs *in vitro* with antisense oligos targeted to the initiation region of nARIA or ARIA mRNA. Then, nARIA or ARIA activity may be assayed in co-culture with LSG or MHN neurons (assay of nAChR macroscopic and single channel 30 currents as above). The efficacy of antisense constructs is confirmed by quantitative RT-PCR of control and antisense-treated explants for nARIA, ARIA and transcripts unaltered by the treatment (e.g. actin). Specificity of the antisense is evaluated by assay of 35 oligomers, equivalent in size and composition but with

20-25% mismatched bases. Presynaptic properties (e.g. electrical activity, transmitter release/mini amplitude) may be tested to control for other non-specific effects of the antisense.

5

Comparison of the affinity of nARIA vs ARIA for heparin sulfate proteoglycan.

MCF7 breast tumor cells were treated with conditioned media from either ARIA or nARIA transiently transfected 10 COS1 cells. Some of the media was prebound with heparin attached to glass beads. Prior to treatment, the beads were pelleted by centrifugation to remove any heparin associated proteins. The supernatant was used to treat the MCF7 cells and tyrosine phosphorylation of the 15 ARIA/nARIA receptor was analyzed. See Figure 15. It appears that the ARIA is binding to heparin and is removed from the media by centrifugation. The nARIA lane shows a slight drop in signal which is unaffected by increasing concentrations of heparin (from 6 μ g - 60 μ g). 20 It is possible that this slight drop is an artifact of differential binding between the lanes and thus nARIA does not bind heparin.

Heparin is a component of both the cellular surface and 25 the extracellular matrix. The difference in the binding affinities of nARIA and ARIA for heparin have two implications. (1) Heparin can affect the affinity of the ligand for the receptor as has been previously shown for HB-EGF. In this case, heparin may increase the affinity 30 for the receptor. Accordingly, a given amount of ARIA may have higher affinity and thus more cellular activity and effect than the equivalent amount of nARIA. This implication may extend to other possible effects of the physiological concentration of the ligand. (2) The lack 35 of affinity for heparin may result in greater solubility

of nARIA *in vivo* since the molecule will not be bound to the extracellular heparin. This possibility may influence the localization of the ligand effects and the point concentration of the ligand.

5

Conclusions

The series of experiments described herein are some examples illustrating possible uses of nARIA. As 10 discussed above herein, there are many other possible therapeutic, diagnostic and pharmacologic uses of nARIA. The biological role of nARIA has been shown to be distinct from that of ARIA and from other members of the heregulin/NDF family. Therefore, nARIA may be useful in 15 therapeutic treatments and as a diagnostic tool for abnormal neuronal conditions. In addition, a comparison of the expression levels and activities of the members of the NDF/hereregulin/ARIA family may prove to be useful in the characterization and treatment of neuronal disorders 20 and abnormal conditions and neurological developmental questions which are at this time unanswered.

Example 4: The n-ARIA isoform of neuregulin is both 25 necessary and sufficient for the induction of acetylcholine-receptors in neurons.

It is clear that the n-ARIA isoform of neuregulin is both necessary and sufficient for the induction of 30 acetylcholine-receptors (nAChR) in neurons. The induction of nAChR expression normally occurs at specific sites within the CNS and PNS during synapse formation and can be mimicked by presynaptic input or presynaptic0input 35 conditioned media. Definitive evidence has been uncovered showing that the expression is mediated by n-

ARIA. The timing and pattern of expression as well as the primary structure and functional effects of n-ARIA differ importantly from the other neuregulins.

5 The expression profile of n-ARIA is distinct from that of the Ig-domain containing forms:

The n-ARIA sequence is unique. An extracellular Ig-like motif, common to all other neuregulins, is replaced by a 10 highly conserved cys rich domain (98% identical chick to human and linked to a β 1 type EGF-like domain.

n-ARIA is the only neuregulin isoform for which expression is restricted to the nervous system (both PCR 15 and Northern analyses).

n-ARIA expression, unlike the Ig containing forms, is apparent at the earliest stages of neuronal differentiation (E2-E3 in chick; PCR and Northern 20 analyses).

The development of isoform specific antibodies and the assessment of the distribution of n-ARIA protein in the CNS and PNS has been performed. A polyclonal sheep 25 antibody was raised which is specific to the cysteine rich domain containing isoforms of the "neu"-regulin gene. This antibody was developed against a peptide sequence encoding the hydrophilic portion within the highly conserved domain. The antibody is capable of recognizing 30 both the denatured and natural protein states on solid matrix support (Figure 16). Immunohistochemical studies demonstrate that this antibody recognizes both avian and mammalian homologues of the nARIA protein. Immunohistochemical studies have also demonstrated that:

Unlike the Ig containing forms, n-ARIA is largely (but not exclusively) expressed by cholinergic and/or cholinoreceptive neurons. n-ARIA expression is prominent in medial septal cholinergic nuclei, basal forebrain, 5 deep cerebellar nuclei, cerebellar Purkinje neurons, and retinal ganglion neurons as well as in cranial, somatic and visceral motor nuclei.

n-ARIA protein appears to be targeted to axons and axon 10 terminals as soon as such projections can be detected either *in vitro* or *in vivo*. n-ARIA immunoreactivity is localized in motor nerve terminal at the neuromuscular junction and "double" staining indicates that n-ARIA expression aligns with (post synaptic) α bungarotoxin 15 binding, which reveals the distribution of muscle AchRs. n-ARIA immunoreactivity is also detected in axonal terminals and *en passant* synapses on CNS and PNS neurons (Figure 17).

20 The functional profile of n-ARIA is distinct from that of the Ig-domain containing forms:

n-ARIA is more soluble than the Ig-domain containing forms and unaffected by heparin and less avidly bound by 25 extracellular matrix proteins.

The activity of n-ARIA in inducing tyrosine phosphorylation in neurons is more potent, more rapid and more persistent than Ig-domain containing forms.

30 The induction of specific nAChR subunits and the enhancement of nAChR-currents by presynaptic input are specifically mimicked by recombinant n-ARIA protein. The activity of n-ARIA in inducing receptor expression in 35 neurons is more potent and more persistent than Ig-domain

containing forms (Figure 18).

The ability to enhance $\alpha 3$ type nAChR subunit gene expression is unique to the n-ARIA isoform.

5

The ability to enhance $\alpha 3$ type nAChR subunit gene expression is unique to the n-ARIA isoform.

Most importantly, the induction of specific nAChR-
10 currents by presynaptic input are selectively blocked by prior exposure of the input to n-ARIA specific antisense-oligonucleotides (Figure 19) or n-ARIA antibodies.

The n-ARIA isoforms

15

The role of n-ARIA in the differentiation and synaptic function of the septal cholinergic neurons that project to hippocampus and amygdala has been investigated. Specifically, evidence suggests that n-ARIA might 20 collaborate with other growth factors, previously implicated in the differentiated function and perhaps, survival of CNS cholinergic neurons.

n-ARIA may induce several parameters of differentiated 25 phenotype in central cholinergic neurons. The effects of n-ARIA \pm target-derived growth factors, on CH synthesis and release have been examined. Transient depletion of n-ARIA (in vitro by antisense or antibody treatments to 30 "knock down" endogenous n-ARIA) has been utilized to initially test for n-ARIA dependent changes cholinergic neurons.

Genomic probes to n-ARIA (i.e. the sequence encoding the crucial cys rich domain) have been developed. The 35 effects of an n-ARIA-selective gene knock out was

examined by using homologous recombination with subsequent excision of the selectable marker employing the Cre-LOXP approach. The n-ARIA exon-specific knock-outs are expected to survive embryogenesis as, unlike 5 other neuregulins (including ARIA), expression is confined to the nervous system, allowing determination of the role of n-ARIA in differentiation, synaptic function and survival of CNS cholinergic neurons.

10 Example: 5

Products of the neuregulin gene and their receptors, the erbBs, (erbB2, 3 and 4) control multiple critical biological processes. Dysregulation of Neuregulin-erbB 15 signaling is associated with major human cancers, specifically in breast, ovarian prostate and lung cancer. In some patients this is accompanied by the appearance of soluble erbBS in the circulation and/or elevated levels of auto-antibodies recognizing the erbBs. 20 If soluble erbBs are predictive of dysregulated erbB-neuregulin expression of function, then sensitive assays for these proteins in patients' blood has potential to provide non-invasive prognostic information on disease status, both in detecting primary disease and in 25 monitoring disease-free status following traditional therapies. Recently we demonstrated that interactions between neuregulin and erbB expressing cells induces membrane to nuclear translocation of the cytoplasmic domain of neuregulin. We constructed a neuregulin-green 30 fluorescent protein fusion protein that allows us to detect erbB-neuregulin interactions. By coupling this assay with fluorescence activated cell sorting, we are devising a sensitive rapid and high throughput assay for the presence of erbBs in biological fluids. A further 35 modification of this assay detects nuclear targeting of

neuregulin-Gal4-regulated reporter gene. The level of reporter gene expression is dependent of the concentration of extracellular stimuli (e.g. erbBs), providing a means of not only detecting soluble erbBs in 5 biological fluids, but quantifying levels. In addition, using reporter gene expression provides an amplification step that significantly increases the sensitivity of the assay.

10 ROLE OF HEN1 in NEUROGENESIS and RECENT DATA on
NEUREGULIN

We found that an embryonic neuronal specific basic-helix-loop-helix protein, HEN1 (also known as NSCL1 or NHLH), 15 interacts with "LIM only" proteins. Examination of the expression patterns of XHEN1 and XLMO-3, the Xenopus homologs of these human genes, reveals extensive overlap during early neurogenesis. Co-expression of these two genes in Xenopus embryos induces a cascade of expression 20 of neuronal specific basic-helix-loops-helix proteins that lead to neuronal differentiation. Recently, I tested whether neuregulin (NRG) cytoplasmic domain might mediate "back signaling". An intense subnuclear 25 localization was observed in cells transfected with the NRG cytoplasmic domain. A similar observation was made in primary neuron immunostained with an antibody against the cytoplasmic tail of NFRG. This nuclear translocation requires a NLS motif at the beginning of the cytoplasmic domain, which includes eight amino acids: KTKKQRKK. One 30 of the functions of this nuclear translocation was to induce apoptosis. Furthermore, a novel gene (CNIP) was found to bind to the cytoplasmic domain of NRG, which might be the modulator for the functions of NRG cytoplasmic domain.

**CNIP: A NOVEL INTERACTOR PROTEIN SPECIFIC FOR THE
CYTOPLASMIC DOMAIN OF CRD NEUREGULIN**

The neuregulin (NRG) gene encodes a number of splice
5 variants that are epidermal growth factor (EGF)-related
polypeptides. The protein structure of most NRG isoforms
includes 3 general domains:

1. The variable extracellular domain, including
the EGF-like domain but differing amongst NRG isoforms by
10 the inclusion of a cysteine rich or an Ig-like domain
(referred to as CRD- or Ig-NRG, respectively,
2. A conserved transmembrane domain and
3. A cytoplasmic domain of unknown functions.

We have begun to explore whether the cytoplasmic domain
15 of CR NRG (also known as nARIA or neuregulin) might
mediate "back signaling" by interaction with specific
cellular proteins. The interaction "hunt" for the
cytoplasmic domain of NRG was performed using a *Xenopus*
cDNA library. cDNAs from the strongest 14 of 4000
20 positives obtained were analyzed. All 14 interactors
encode the same product. To test the specificity of the
putative interactor, a panel of control baits were
screened for binding with the gene product: only lexA-
CRD-NRG cytoplasmic domain interacted strongly.
25 Interactions with the cytoplasmic domains of TGF β
receptor, G proteins, cyclin, cyclin dependent kinase or
Myc proteins do not occur. The expression pattern of the
CRD-NRG Interactor Protein (CNIP was examined in *Xenopus*
and found to be strikingly similar to that of CRD-NRG.
30 Thus CNIP is prominently expressed in the nervous system,
especially in the hindbrain, eyes and spinal cord. In
view of the specificity of the interactions as well as
their overlapping patterns of expression with the nervous
system CNIP-NRG interactions may occur *in vivo*.

NOVEL FUNCTIONS OF THE CYTOPLASMIC DOMAIN OF NEUREGULIN

Neuregulins (NRGs) comprise a large family of EGF-like growth factors expressed in both the CNS and PNS. The 5 NRG 1 gene encodes multiple splice variants including secreted and transmembrane isoforms. The external (N-terminal) portion of both membrane anchored and secreted NRG isoforms includes a characteristic Ig-like or cysteine rich domain, and an EGF-like domain that is 10 essential for NRG-ErbB interactions. Membrane anchored and secreted NRG isoforms identified to date include one of three distinct (a, b or c-type) cytoplasmic domains. Although the cytoplasmic domains are highly conserved (85% identity from chick to human), the biological 15 function is unknown.

We tested whether NRG cytoplasmic domain(s) might mediate "back signaling" in NRG expressing cells, following interaction of the tethered ligands with erbB receptors. 20 And intense subnuclear localization was observed in cell transfected with the NRG cytoplasmic domain. A similar observation was made in primary neuron immunostained with an antibody against the cytoplasmic tail of NRG. This nuclear translocation requires a NLS motif at the 25 beginning of the cytoplasmic domain, which includes eight amino acids: KTKKQRKK. The motif is highly conserved (the same sequence for human, rat, mouse, chick and Xenopus NRG). The nuclear translocation was augmented by activation of PKC, or cell expressing erbB receptors. 30 One of the functions of this nuclear translocation was to induce apoptosis. Furthermore, a novel gene (CNIP) was found to bind to the cytoplasmic domain of NRG, which might be the modulator for the functions of NRG cytoplasmic domain.

CRD-NRG IN MOUSE PERIPHERAL NERVOUS SYSTEM DEVELOPMENT.

Synaptogenesis at nerve-muscle junctions involves the redistribution of preexisting surface acetylcholine receptors (AchRs), as well as increased local synthesis and insertion of new receptors. The latter increase in nAChR expression is induced by members of the neuregulin (NRG) family (Fischbach and colleagues *Cell*, 72:801;1993). We have characterized herein an NRG isoform in chicks, mice and humans in which a highly conserved cysteine-rich domain replaces the Ig-motif found in other NRGs; we have termed this variant CRD-NRG. CRD-NRG appears to be both a necessary and sufficient signal for the control of neuronal nAChR expression during synaptogenesis (*Neuron*, 20:255;1998). In situ hybridization analysis in embryonic mice reveals that CRD-NRG is highly expressed in all cranial nerve nuclei, visceral and somatic motor neurons, and in numerous CNS regions, including the olfactory and vomeronasal sensory neurons, the main and accessory olfactory and vomeronasal sensory neurons, the main and accessory olfactory bulbs, hippocampus, amygdala, thalamus, and hypothalamus, as well as septal cholinergic and dopaminergic nuclei, we now evaluate the role of NRG1 signaling in the development of the peripheral nervous system using CRD-NRG specific knockout mice by in situ, immunocytochemical, and immunohistochemical studies.

Neuregulins, originally referred to as ARIA, heregulin (HRG), neu differentiation factor (NDF), glial growth factor (GGF), and sensorimotor-derived factor (SMDF), constitute a large family of structurally related glycoproteins which are produced as a consequence of alternative splicing of the *neuregulin-1* (NRG-1) gene.

Neuregulins function as ligands for the erbB family of receptor tyrosine kinases. Targeted disruptions of the entire NRG-1 gene, and of the genes for its three receptors, erb2, erb3, erb 4 have been generated
5 (Gassmann et al., 1995, Lee et al. 1995, Meyer and Birchmeter, 1995, Kramer et al. 1996, Riethcacher et al. 1997, and Erickson et al., 1997). The phenotype of these mice demonstrated the multiple essential roles of this
10 signaling system in the formation of the peripheral nervous system and the heart.

As many as 14 different neuregulin cDNAs have been isolated (Fischback and Rose, 1997). While all known NRG isoforms contain an EGF-like repeat which is considered
15 essential for NRG-1 activity, isoforms may or may not be have a kringle domain, an Ig-like domain, a spacer domain, a transmembrane domain, and/or a variety of different cytoplasmic tails. However, all isoforms can be broadly classified into two currently mutually
20 exclusive categories: isoforms which contain an If-like domain N-terminal to the EGF-like domain (Ig-NRGs), such as ARIA, HRG, NDF, and Ggf, and isoforms which contain a cysteine-rich domain N-terminal to the EGF-like domain (CRD-NRFs), sucha s SMDF.

25 Different classes of isoforms show distinct patterns of spatial and temporal expression during embryogenesis (Meyer et al. 1997), suggesting that the different isoforms could be mediating distinct biological signals.
30 Analysis of mice null for Ig-containing NRGs and comparative analysis of mice null for all NRGs or just Ig-containing NRFs provides further evidence to support this conclusion (Meyer et al 1997, Sandrock et al. 1997)

35 We have found by in situ hybridization analysis in

embryonic mice that CRD-NrG is highly expressed in all cranial nerve nuclei, visceral and somatic motor neurons, and in numerous CNS regions, including the olfactory and vomeronasal sensory organs, the main and accessory 5 olfactory bulbs, hippocampus, amygdala, thalamus, and hypothalamus, as well as septal cholinergic and dopaminergic nuclei. In addition, we have generated mice which are homozygous null exclusively for all CRD-containing NRG-1 isoforms. We now present evidence for 10 distinct and essential roles of CRD-mediated NRG-1 signaling in the development of the peripheral and central nervous systems.

15

Example 6:

Neuregulins constitute a large family of structurally related glycoproteins produced by alternative splicing of 20 the *neuregulin-1* gene.^{1,2} Neuregulin signaling, mediated by activation of the erbB family of receptor tyrosine kinases, has been implicated in the inductive interactions between pre- and post-synaptic partners at developing nerve-muscle and nerve-nerve synapses.^{1,3-5} 25 Gene targeting to selectively disrupt cysteine rich domain (CRD) containing NRG-1 isoforms in mice reveals that CRD-NRG-1 is required for synaptogenesis in the peripheral nervous system. In CRD-NRG-1^(-/-), mice, peripheral projections defasciculate and display aberrant 30 branching patterns upon arrival within their targets. The profuse sprouts of motor projections are transiently associated with broad bands of postsynaptic ACh receptor clusters. Schwann cell precursors, that initially accompany peripheral projections, are absent from axonal 35 terminations within muscle. Following an aborted attempt

at synapse formation, sensory and motor nerves withdraw and eventually degenerate. Our data demonstrate the essential role of CRD-NRG-1 mediated signaling for coordinating nerve, target, and Schwann cell interactions 5 in the formation and maintenance of peripheral synapses.

Alternative splicing generates at least fifteen different *neuregulin-1* (NRG-1) cDNAs.^{1,6} All identified NRG-1 isoforms can be broadly classified into three mutually exclusive 10 categories: Type I and II isoforms that contain an Ig-like domain (Ig-NRGs) and Type III isoforms that contain a cysteine-rich domain (CRD-NRGs) N-terminal to a common EGF-like sequence (Figure 20A).^{3,6-11} Although different NRG-1 isoforms show distinct patterns of spatial and 15 temporal expression during embryogenesis (data not shown), it is not clear to what extent these isoforms mediate distinct biological responses.^{3,11} Mice homozygous for disruptions of all NRG-1 isoforms, all Ig-NRG-1 isoforms, or all isoforms containing a cytoplasmic tail 20 die at E10.5 from heart defects, prior to significant expression of CRD-NRG-1 isoforms.^{12,15} As such, these mice provide no direct evidence as to the possible functions of CRD-NRG-1 isoforms in neural development or synaptogenesis.

25 CRD-NRG-1 isoforms are the predominant NRG-1 transcripts in the murine nervous system after E10.5. In particular, the temporal pattern of CRD-NRG-1 expression in hindbrain and spinal cord visceral and somatic motor neurons, as 30 well as in cranial and trunk sensory ganglia, suggests an important role for this isoform in synapse formation (Figure 20E. b & data not shown).^{3,5,10,11} To assess the potential role of CRD-NRG-1 isoforms in neural development, we generated mice selectively lacking all 35 NRG-1 isoforms that contain a CRD domain within the N-

terminus.

Experimental Method

5 CRD-NRG-1 ^(+/-) embryonic stem (ES) cells were isolated following homologous recombination with a mutated CRD-encoding exon that included a nonsense mutation/frameshift and a novel XbaI site inserted into the codon immediately 3' of the last methionine (Figure 10 1B). These ES cells were used to generate germline chimeras following injection into mouse blastocysts. Male chimeras transmitting the mutant allele were used to produce fertile heterozygote offspring.

15 CRD-NRG-1 ^(+/-) mice were interbred to produce homozygote CRD-NRG-1 ^(-/-) mice. In the resulting litters, cleaned, dead pups were consistently found among viable, healthy littermates (Figure 20E. a). Genotype analysis identified the dead pups as CRD- NRG-1 ^(-/-) (22% of first 132 PO pups) 20 (Figure 20C). As expected, mutant animals expressed both Ig-containing and mutant CRD-containing mRNAs (Figure 20D). *In situ* hybridization revealed that the spatial and temporal expression pattern of the nonsense mutation containing-CRD-NRG-1 mRNA was not altered in the mutants 25 (Figure 20E, b vs. b¹, & data not shown).

CRD-NRG-1 ^(-/-) pups were limp, and limb movement was not detectable (Figure 20E,a). These pups did not breathe, rapidly became cyanotic, and finally were unresponsive to 30 all stimuli despite the presence of a heartbeat for several minutes after birth. Histological analysis confirmed that hearts of CRD-NRG-1 ^(-/-) mice were normal but that the lung alveoli failed to expand, confirming that CRD-NRG-1 ^(-/-) pups never take a breath (data not 35 shown and Figure 20E. c vs. c').

Examination of PO mutant diaphragms revealed only remnants of nerve-muscle synapses (Figure 21A, a vs. a'). Mutants had decreased skeletal muscle mass, and mutant skeletal muscle had central nuclei with prominent 5 nucleoli compared to the elongated, eccentric, peripheral nuclei in control muscle (Figure 21A, d vs. d' and data not shown). CRD-NRG-1^(-/-) mice lacked an intra-diaphragmatic phrenic nerve plexus (Figure 21A, a vs. a'). Bands of α -bungarotoxin (α BgTx) positive 10 acetylcholine receptor (AChR) clusters, reminiscent of end-plate zones, were present but abnormally broad (~6 times wider than littermate controls) (Figure 21A. b vs. b). These diffuse bands of AChR clusters, the absence of associated nerve, and the scattered neurofilament(+) 15 fragments indicate prior presence, and subsequent degeneration, of motor projections. Indeed, examination of diaphragms from E14.5, E15.5, E18.5, and PO mutant mice revealed elaborate phrenic nerve arborization at E14.5 and progressive loss of these nerve fibers during 20 subsequent stages of development (Figure 21B, f vs. f, and data not shown). Phrenic nerve processes in E14.5 CRD-NRG-1^(-/-) mice were grossly abnormal with extensive defasciculation, aberrant axonal branching patterns, and projections extending as far as the most lateral borders 25 of the diaphragm (Figure 21B, f vs. f'). Although post-synaptic AChR clusters were sometimes detected subjacent to neurofilament(+) and synaptophysin(+) structures in mutant E14.5 diaphragms, these clusters were typically more diffuse and more often devoid of associated nerve 30 than age-matched controls (Figure 21B, g vs g').

A similar pattern of defective nerve-muscle interaction was detected in intercostal and limb muscle groups (e.g., scattered α BgTx(+) - AChR clusters and lack of co- 35 localization with nerve; Fig 2B, e vs. e' and data not

shown). Acetylcholinesterase (AChE), which is normally enriched at the site of nerve-muscle contact following the establishment of synapses, was scattered and faint in mutant muscle (Figure 21A, c vs. c').¹⁶ Thus, in CRD-NRG-1⁵ mice, motor axons initially extend to and contact the target muscle and some aspects of pre- and post-synaptic specializations were evident. However, as these preliminary synaptic interactions were not sustained, we next examined if the number of motor neurons was 10 affected.

Somatic motor neurons were visualized by *in situ* hybridization to the vesicular ACh transporter (VAT) mRNA.¹⁷ The number of VAT(+) cell bodies within the 15 ventral horn of C1-C6 of E18.5 mutants was 60% less than that of controls (Figure 22A. a vs. a', b). A dramatic reduction in the diameter of peripheral nerves arising from motor pools at other spinal cord levels, including those within intercostal and hindlimb muscles, was also 20 evident in E18.5 CRD-NRG-1^(-/-) animals compared to controls (Figure 21B. e vs. e' and data not shown). Likewise. E18.5. but not E14.5. cervical dorsal root ganglia and their peripheral sensory projections were dramatically decreased in size in CRD-NRG-1^(-/-) compared 25 with control animals (Figure 22B, d vs. d' and data not shown). The observed degeneration of peripheral sensory and motor nerve projections by E18.5 is likely to account for the abnormal morphology of forelimbs and absence of all limb movement, evident in both E18.5 and PO CRD-NRG-1 30^(-/-) mice (Figure 20E, a and data not shown).

We next examined the development of the sensory and motor projections prior to arrival in their respective target fields. At E1 1, whole mount immunostaining of mutant 35 embryos with β -Tubulin III, a pan-neuronal marker,

revealed that initial projections of both sensory and motor neurons were fasciculated (Figure 23B, h vs. h').¹⁸ At later stages and at more caudal levels of mutant embryos, these terminal projections displayed dramatic 5 defasciculation, aberrant and irregular branching, and markedly increased terminal sprouting (Figure 22B, f vs. f'). Likewise, in caudal hindlimb muscle of older mutant animals, supernumerary terminal motor sprouts were visualized with VAT Ab staining (Figure 22B, g vs. g'). 10 The VAT(+) sprouts and AChR clusters, visualized as α BgTx "hot spots," were each detected without the complementary marker of nerve-muscle synapse formation (data not shown). Finally, at later times and at more rostral levels of mutant embryos, the terminal projections of 15 sensory and motor neurons were smaller, bulb-shaped, and compact, when compared to the more splayed nerve endings in control animals (Figure 22A, c vs c': 22B, e vs e').

Taken together, these data indicate a temporal and 20 rostral-caudal pattern whereby the initial trajectory and outgrowth of peripheral nerves in CRD-NRG-1 ^(-/-) mice are grossly normal. However, once within the terminal fields, projections defasciculate, displaying profuse and aberrant branching, and subsequently, appear bulbous and 25 then fragmented, consistent with their ultimate retraction and degeneration.

Peripheral nerves projecting to their targets are lined by neural crest-derived Schwann cell precursors 30 that express erbBS, p75 NGF receptor, and S100 β .^{19,20,21} At E18.5, mutant embryos lacked erbB3/p75-expressing cells in both dorsal and ventral roots (Figure 22B, d vs. d' and data not shown). Likewise, S100 β expressing cells were absent from intramuscular branches of the phrenic 35 nerve in E14.5 mutant diaphragms (Figure 2B, h vs h').

Although erbB3(+) cells were detected along nerve roots in E11 mutant mice, the progressive, rostral-caudal loss of these cells was evident by E12 (Figure3B, h vs. h'; Figure 4B, j vs. j'). The spatial pattern of Schwann cell 5 precursor loss was revealed by examination of more caudal structures in older CRD-NRG-1^(-/-) animals. In peripheral hindlimb muscle of E18.5 CRD-NRG-1^(-/-) mice, S100 β (+) cells lined the VAT(+) axons and preterminal branches but were not detected at the site of VAT (+) nerve 10 terminations (Figure 22B, I) In contrast, in control embryos VAT(+) preterminal nerve branches and nerve terminations were always associated with S100p (+) staining (Figure 22B, I¹).

Cranial sensory ganglia strongly express CRD-NRG-1 15 isoforms (Figure 20E. b: Figure23A, a, b, and data not shown)." CRD-NRG-1. erbBS and erbB4 mRNA have been detected in the peripheral targets of these ganglia, including vibrissa follicles (Fig 23A, c-e).¹¹ The fate of these neuronal populations and their associated erbB3(+) 20 Schwann cells was significantly altered in CRD-NRG-1^(-/-) mice. Both neural crest and otic placode-derived cellular elements of cranial ganglia appear to form in CRD-NRG-1^(-/-) mice, but their initial projections were markedly less robust in mutants (Figure 23B, h vs. h'). As cranial 25 nerves enter their target fields, the pattern of nerve defasciculation and extensive branching was seen (Figure 23B, h vs. h'). The subsequent demise of cranial nerve projections was apparent by E12.5, and by E14.5. cranial ganglia (including the trigeminal and geniculate) and 30 their associated nerves were dramatically reduced in size in CRD-NRG-1^(-/-) mice (Figure 23B. i vs. i' and data not shown). Furthermore, cranial nerve projections were nearly devoid of associated Schwann cell precursors in CRD-NRG-1^(-/-) mice compared with controls (Figure 23B, i 35 vs. i' and data not shown). Examination of cranial nerves

at earlier stages of development reveals the initial appearance and subsequent loss of erbB3(+) cells along the ascending spinal accessory (N11n) nerve and the coalescence of vagus nerve (N10n) and in the ganglia and 5 along the peripheral projections of the trigeminal (N5g), geniculate (N7g), glossopharyngeal (N9g), and vagal N10g) ganglia (Figure 23B, j vs. j and data not shown).

CRD-NRG-1 is expressed in VAT(+) cells of cranial motor 10 nuclei, such as the trigeminal (N5N), facial (N7N), and dorsal motor nucleus of the vagus (Figure 23A, f & g and data not shown). At E18.5 the number and distribution of VAT(+) neurons within the brainstem of CRD-NRG-1 ^(-/-) embryos was abnormal with the most striking changes seen 15 in the motor nuclei of cranial nerves N5N and N7N (Figure 4B, k vs. k'; i vs. i'). Preliminary analysis of other VAT(+) neurons within brainstem and forebrain nuclei also indicates possible changes in development and synaptogenesis (D.W. and L.W.R., in preparation). 20 In summary, CRD-NRG-1 is an essential, nerve-derived component of peripheral synaptogenesis. The phenotype associated with the, selective disruption of CRD-NRG-1 mediated signaling is distinct from all other NRG-1 ligand or receptor knockouts reported to date. CRD-NRG-1 25 ^(-/-) mice die within minutes of birth, unable to breathe due to the lack of functional phrenic nerve-diaphragm synapses. The defects in CRD-NRG-1 ^(-/-) mice are evident as a temporal progression and in a rostral-caudal pattern during embryogenesis. Thus, both motor and sensory nerve 30 projections initially emanate from hindbrain to lumbar spinal cord levels. However, with the arrival of projections within their respective targets, mutant peripheral nerves branch extensively, and Schwann cell precursors, initially associated with peripheral 35 projections, do not survive. By parturition, CRD-NRG-I ^(-/-)

’ mice have severe deficits in the number and extent of motor and sensory neurons, consistent with extensive neural degeneration. At birth, skeletal muscle targets display features of immaturity similar to those found in 5 human myotubular myopathy.²² Genetic defects in a muscle phosphatase have been implicated in the severe X-linked form of this disease.²³

NRG-1 signaling has long been implicated as an important 10 component for survival and differentiation during Schwann cell development.^{2,13,19} Loss of Schwann cell precursors is a phenotype shared by CRD-NRG-1 ^(-/-) and erbBS null mice.²¹ The early effects of postnatal axotomy on Schwann cell apoptosis are partially reversed by exogenous NRG-1 15 protein.^{20,24} Furthermore, previous studies comparing NRG-1 ^(-/-) mice and Ig-NRG-1 ^(-/-) mice led to the proposal that non-Ig containing NRG isoforms might be required for Schwann cell survival.¹¹ The unique neural specificity of CRD-NRG-1 expression in mouse and the progressive loss of 20 erbB3(+) Schwann cell precursors along peripheral nerve projections in CRD-NRG-1 ^(-/-) mice implicate CRD-NRG-1 isoforms as the essential, nerve-derived trophic signal for erbBS-mediated Schwann cell survival.

25 Sensory and motor neurons are critically dependent on target-derived trophic support during early development^{25,26} Likewise, the development and terminal differentiation of target muscle requires interaction with motor nerve. The phenotype of CRD-NRG-1 ^(-/-) mice 30 demonstrates that CRD-NRG-1 isoforms are necessary components comprising the reciprocal cascades of these cellular interactions. Birth, initial differentiation and peripheral projections of motor and sensory neurons proceed apace in the absence of CRD-NRG-1 isoforms. 35 Initial aspects of post-synaptic organization, such as

clustering of AChRs in muscle, are also detected in CRD-NRG-1 ^(-/-) mice. The AChR-cluster inducing signal, agrin, acts through MuSK receptor complexes in skeletal muscle.^{29,30} It is noteworthy that mice in which these 5 genes are disrupted show abnormalities in neuromuscular junction formation, but motor projections neither withdraw nor degenerate.^{29,30}

In contrast, the genetic disruption of CRD-NRG-1 results 10 in both pre- and post-synaptic defects consistent with loss of reciprocal, inter-dependent nerve and target-derived trophic support. Furthermore, the target-derived support that appears to be required for the persistence 15 of the incoming nerves is not constitutively available but must be elicited by a nerve-derived signal. We propose that this nerve-derived signal is CRD-NRG-1. As Schwann cell precursors might also provide essential trophic support for developing nerves, and these cells fail to survive in CRD-NRG-1 ^(-/-) mice, it is possible that 20 Schwann cell precursors and/or peripheral targets respond individually or synergistically to a common nerve-derived, CRD-NRG-1-mediated signal that normally elicits context-dependent, trophic and synaptogenesis.

25 **METHODS**

Generation of CRD-/- Mice. A mouse 129/SV genomic library (Lamba EMBL3, Stratagene) was screened with a mouse CRD-specific probe. Positive clones were mapped and sequenced 30 revealing that the CRD-domain is encoded by a single exon. PCR-mediated site-directed mutagenesis and standard protocols in molecular biology were used to create the targeting vector. ES cells were electroporated, selected for G418 resistance (150 µg/ml active substance), and 35 screened by Southern blot for homologous recombination.

ES clones, in which homologous recombination resulted in replacement of one of the wild-type NRG-1 alleles with the mutant gene, were injected in C57/BL6/J blastocysts. Chimeric males were mated with 129/Sv females to generate 5 heterozygotes on a pure 129/Sv background.

RT-PCR. Total brain/spinal cord RNA was isolated using Trizol (Gibco BRL) following the manufacturers protocol, and first-strand cDNA was made using Superscript II 10 reverse transcriptase following the manufacturers protocol (Gibco BRL). Mouse-specific primers and oligonucleotide probes corresponded to the following: from Genbank L41827, CRD 5' primer (549-565), CRD-probe (582-605); from Genbank u02318: Ig 5' primer (499-518), 15 Ig/CRD 3' primer (885-903), egf-cyt primers (913-931), (1184-1203), Ig probe (608-631), TM probe (1080-1103). Note thgt all primer pairs span exon-intron boundaries.

In Situ Hybridization and Cell Counts. In situ hybridization (ISH) probes were as follows: mouse probes 20 specific for CRD domain (corresponding to Genbank AF045654, 607-1206), PAN-NRG-1 (corresponding to Genbank AF045654, 867-1277), erbB3 (Corresponding to Genbank u29339, 3321-4113), erbB4 (corresponding to Genbank 25 L07868, 3088-3957), VAT (gift of Dr. J. Dedman; Genbank AF019045, 945-2157) or rat 75 NGF receptor (gift of Dr. Moses Chao). Embryos were fixed in 4% paraformaldehyde by transcardial perfusion, followed by post-fixation overnight at 4°C. Following cryoprotection in 30% 30 sucrose, embryos were embedded in OCT and 12 µm sections were cut on a cryostat. ISH was done as previously described.³¹ For cell counts, 10 µm transverse serial sections were cut through C1-C6 and stained for VAT. Total number of nuclei of VAT(+) cells in every sixth 35 section was then counted. Counts between ages are not

strictly comparable as total cell numbers were not corrected for split or multiple nucleoli. We feel our counts reflect a conservative estimate of the cell loss, as motor neurons with retracted processes are included.

5 Whole-mount ISH was done as previously described.³²

Histological Analysis and Immunostaining. Lung was fixed overnight in 4% paraformaldehyde and embedded in paraffin. 8 μ m sections were cut and stained with H&E. Diaphragm was sandwiched between liver, flash-frozen in 10 liquid-nitrogen cooled isopentane, and 6 μ m sections were stained with H&E. AChE staining was done as described previously.³³ Whole-mount immunostaining using monoclonal antibody to β -Tubulin III (Sigma) was done as previously described.³⁴ After staining, embryos were dehydrated and 15 cleared in cedarwood oil. Whole-mount diaphragm staining was done as previously described.^{29,30} Immunofluorescence labeling employed monoclonal anti-neurofilament NF-60 & 168 antibodies (Sigma), anti-S100 β antibody (DAKO), FITC-conjugated alpha btx (Molecular Probes), anti-20 synaptophysin antibody (gift of Dr. Pietro de Camilli), and rhodamine or FITC-conjugated anti-mouse or anti-rabbit antibodies (Jackson Immunoresearch).

References

References to Examples 1-5

5 Abuchowski et al., In: "Enzymes as Drugs", Holcnenberg et al., eds. Wiley-Interscience, New York, NY, 367-383 (1981).

10 Arenella, L.S., Oliva, J.M. Jacob, M. H. (1993) Reduced levels of acetylcholine receptor expression in chick ciliary ganglion neurons developing in the absence of innervation. *J. Neuroscience* 13(10):4525-37.

15 Ausubel, F., Brent, R., Kingston, R., Moore, D., Seidman, J.G., Smith, J. and Struhl, K. eds. (1994) Current protocols in Molecular Biology Green Publishing Associates, Inc.

20 Berg, D.K., Boyd, R.T., Halvorsen, S.W. Higgins, L.S., Jacob, M.H., Margiotta, J.F. (1989) Regulating the number and function of neuronal acetylcholine receptors. *Trends Neurosci.* 12:16-21.

25 Bertrand, D., Cooper, S., Valera, D. and Ballivet, M. (1991) Electrophysiology of neuronal nicotinic acetylcholine receptors expressed in *Xenopus* oocytes following nuclear injection of genes or cDNAs. *Med Neurosci.* 4:174-193.

30 Betz, H. (1990) Homology and analogy in transmembrane channel design: lessons from synaptic membrane proteins. *Biochemistry* 29:3591-99.

35 Boulter, J., Evans, K., Goldman, D., Martin, G., Treco., D., Heinemann, S., Patrick, J. (1986) Isolation of a cDNA

clone coding for a possible neural nicotinic acetylcholine receptor -subunit. *Nature* 319:368-74.

Boyd, R.T., Jacob, M.H., Couturier, S., Ballivet, M.,
5 Berg, K.D. (1988) Expression and regulation of neuronal acetylcholine receptor mRNA in chick ciliary ganglia. *Neuron* 1:495-502.

Brussaard, A.B., Yang, X., Doyle, J.P., Huck, S., Role, L.W. (1994) Developmental regulation of multiple nicotinic ADhR channel subtypes in embryonic chick habenula neurons: Contributions of both the 2 and 4 subunit genes. *Pflugers Archiv.* (In press.).

10 Brussaard, A.B., McGehee, D.S. and Role, L.W. (1994b) Presynaptic input, in the absence of target contact, recapitulates a subset of the modifications of nAChR channels induced *in vivo* to enhance synaptic transmission. (to be submitted to *J. Physiol.*)

20 Carpenter et al., *Toxicol. Appl. Pharmacol.*, 18:35- 40 (1971).

Clarke, P.B.S., Hamill, G.S., Nadi, N.S., Jacobowitz, D.M., Pert, A. (1986) ³H-nicotine-and ¹²⁵I-alpha-bungarotoxin-labeled nicotinic receptors in the interpeduncular nucleus of rats. II. Effects of habenular deafferentation. *J. Comp. Neur.* 251:407-13.

25 Conroy, W.G., Vernallis, A.B., Berg, D.K., (1992) The 5 gene product assembles with multiple acetylcholine receptor subunits to form distinctive receptor subtypes in brain. *Neuron* 9:679-91.

30 Deneris, E.S., Connolly, J., Rogers, S.W., Duviosin, R.

(1991) Pharmacological and functional diversity of neuronal nicotinic acetylcholine receptors. *Trends Pharmacol. Sci* 12:34-40.

5 Devay, P., Qu, X. and Role, L.W. (1994) Developmental regulation of nAChR subunit gene expression during establishment of pre- and post-synaptic connections of lumbar sympathetic neurons of embryonic chicken. *Devel. Biol.* 162:56-70.

10 Engisch, K.L., Fischbach, G.D. (1994) The development of ACH- and GABA-activated currents in embryonic chick ciliary ganglion neurons in the absence of innervation *in vivo*. *J. Neurosci.* 12:1115-25.

15 Falls, D., Rosen, K., Corfas, G., Lane, W. and Fischbach, G.D (1993) ARIA, a protein that stimulates acetylcholine receptor synthesis, is a member of the Neu ligand family. *Cell* 72:801-815.

20 Fischbach, G.D., Role, L.W. and Hume, R.I. (1984) The accumulation of acetylcholine receptors at nerve- muscle synapses in culture. In: *Cellular and Molecular Biology of Neuronal Development*. I. Black (ed.), Plenum Press. pp 25 107-115.

25 Gardette, R., Listerud, M.D., Brussaard, A.B., Role, L.W. (1991) Developmental changes in transmitter sensitivity and synaptic transmission in embryonic chicken sympathetic neurons innervated *in vitro*. *Dev. Biol.* 147:83-95.

30 Gryniewicz, G., Poenie, M. and Tsien, R.Y. (1985) A new generation of Ca^{2+} indicators with greatly improved 35 fluorescence properties, *J. Biol. Chem.* 260:3440-3450.

Habecker, B. and Landis, S. (1994) Noradrenergic Regulation of Cholinergic Differentiation. *Science* 264:1602-1604.

5 Hammil, O.P., Marty, A., Neher, E., Sakmann, B. and Sigworth, F.J. (1981) Improved patch-clamp techniques for high-resolution current recording from cells and cell-free membrane patches. *Pflugers Arch.* 391:85-100.

10 Hasselmo, M.E. and Bower, J.M. (1993) Acetylcholine and memory. *Trends in Neurosci.* 16:218-222.

Heinemann, S., Boulter, J., Deneris, E., Connolly, J., Duvousin, R., Papke, R., Patrick, J. (1990) The brain
15 nicotinic acetylcholine receptor gene family. *Progr. Brain Res.* 86:195-203.

Holmes, W.E., Sliwkowski, M.X., Akita, R.W., Henzel, W.J., Lee, J., Park, J.W., Yansura, D., Abadi, N., Raab, H., Lewis, G.D., et al. (1992) Identification of heregulin, a specific activator of p185erbB2. *Science* 256:1205-1210.

20 Jacob, M.H. (1991) Acetylcholine receptor expression in developing chick ciliary ganglion neurons. *J. Neurosci.* 11:1701-12.

25 Katre et al., *Proc. Natl. Acad. Sci. USA* 84:1487- 1491 (1987).

30 Köhler and Milstein *Eur. J. Immunol.* 6: 511-519 (1976).

Kriegler, M. (1990) *Gene Transfer and Expression: A Laboratory Manual*, Stockton Press, New York, New York.

Levey, M.S., Brumwell, C., Dryer, S., Jacob, M. (1994) Innervation and target tissue interactions differentially regulate acetylcholine receptor subunit transcript levels in Lipscombe, D., Madison, D.V., Poenie, M., Reuter, H.,

5 Tsien, R.W. and Tsien, R.Y. (1988) Imaging of cytosolic Ca²⁺ transients arising from Ca²⁺ stores and Ca²⁺ channels in sympathetic neurons. *Neuron* 1:355-365.

Listerud, M., Brussard, A.B., Devay, P., Colman, D.R.,

10 Role, L.W. (1991) Functional contribution of neuronal AChR subunits by antisense oligonucleotides. *Science* 254:1518-21.

Marchionni, M.A., Goodearl, A.D., Chen, M.S., Birmingham, M.O., Kirk, C., Hendricks, M., Danehy, F., Misumi, D., Sudhalter, J., Kobayashi, K., et al., (1993) Glial growth factors are alternatively spliced erbB2 ligands expressed in the nervous system. *Nature* 362:312- 318.

20 Mandelzys, A., Pie, B., Deneris, E.S., Cooper, E. (1994) The developmental increase in ACh current densities on rat sympathetic neurons correlates with changes in nicotinic Ach receptor α -subunit gene expression and occurs independently of innervation. *J. Neurosci.*

25 15:2357-64.

Margiotta, J.F., Gurantz, D. (1989) Changes in the number, function and regulation of nicotinic acetylcholine receptors during neuronal development. *Dev. Biol.* 135:326-39.

30

McGehee, D.S., and Role, L.W. (1995) Physiological Diversity of nicotinic acetylcholine receptors expressed by vertebrate neurons. *Annual Review of Physiology* (in 35 press).

McGehee, D.S., Yang, X., Devay, P., Heath, M.J.S., Role, L.W. Nicotine potentiates synaptic transmission through presynaptic aBgTx sensitive acetylcholine receptors. *Soc. Neurosci. Abstr.* 19:463.

5 Moss, B.L. and Role, L.W. (1993) Enhanced ACh sensitivity is accompanied by changes in ACh receptor channel properties and segregation of ACh receptor subtypes on sympathetic neurons during innervation *in vivo*. *J. Neurosci.* 13:13-28.

10 Moss, B.L., Schuetze, S.M., Role, L.W. (1989) Functional properties and developmental regulation of nicotinic acetylcholine receptors on embryonic chicken sympathetic neurons. *Neuron* 3:597-607.

15 Nef, P., Oneyser, S., Alliod, D., Couturier, S., Ballivet, M. (1988) Genes expressed in the brain define three distinct neuronal nicotinic acetylcholine 3-
20 receptors. *EMBO J.* 7:595-601.

Newmark et al., *J. Appl. Biochem.* 4:185-189 (1982).

Press, W., Flannery, B., Teukolsky, S. and Vettering, W.P. (1986) Are two distributions different? Komolgorov-Smirnov test. In *Numerical Recipes: The Art of Scientific Computing* (New York: Cambridge Univ. Press) pp 472-475.

30 Ramirez-Latorre, J., Qu, Z., and Role, L.W. (1993) Participation of 5 in neuronal nicotinic AChR channels. *Soc. Neurosci. Abstr.* 19:1533.

35 Role, L.W. (1988) Neural regulation of acetylcholine sensitivity of embryonic sympathetic neurons. *Proc. Natl.*

Acad. Sci. USA 85:2825-2829.

Role, L.W. (1992) Diversity in primary structure and function of neuronal nicotinic acetylcholine receptor channels. *Curr. Opin. Neurobiol.* 2:254-62.

10 Sambrook, J., Fritsch, E.F. and Maniatis, T. (1989). Molecular cloning: a laboratory manual (Cold Spring Harbor Laboratory Press, Cold Spring Harbor, NY), 2nd Ed.

Sargent, P.B. (1993) The diversity of neuronal nicotinic acetylcholine receptors. *Ann Rev. Neurosci.* 16:403-33.

15 Seguela, P., Wadiche, J., Dineley-Miller, K., Dani, J.A., Patrick, J.W., (1993) Molecular cloning, functional properties and distribution of rat brain 7: a nicotinic cation channel highly permeable to calcium. *J. Neurosci.* 13:596-604.

20 Sendecor and Cochran, (1989)

Simmons, L.K., Moss, B.L., Schuetze, S.M. and Role, L.W. (1988) Developmental regulation and modulation of neuronal nicotinic acetylcholine receptor channels in 25 Nicotinic Acetylcholine Receptors in the Nervous System. NATO ASI series H: Cell Biology. Vol. 25H, Ed. F. Clementi, C. Gotti & E Sher, Springer-Verlag, Berlin-Heidelberg, pp 379-392.

30 Smith, M.A., Margiotta, J.F., Berg, D.K. (1983) Differential regulation of acetylcholine sensitivity and -bungarotoxin-binding sites on ciliary ganglion neurons in cell culture. *J. Neurosci.* 7:149-70.

35 Tsien, R.Y. (1989) Fluorescent probes of cell signalling.

Ann. Rev. Neurosci. 12:227-253.

Vernallis, A.B., Conroy, W.g., Berg, D.K. (1993) Neurons assemble acetylcholine receptors with as many as three 5 kinds of subunits while maintaining subunit segregation among receptor subtypes. Neuron. 10:451-64.

Wada, E., Wada, K., Boulter, J., Deneris, E., Heinemann, S., Patrick, J., Swanson, L.W. (1989) Distribution of 2, 10 3, 4, and β 2 neuronal nicotinic receptor subunit mRNAs in the central nervous system: a hybridization histochemical study in the rat. J. Comp. Neurol. 284:314:35.

15 Wen, D., Peles, E., Cupples, R., Suggs, S.V., Bacus, S.S., Luo, Y., Trail, G., Hu, S. Silbiger, S.M., Ben Levy, R. et al., (1992). New differentiation factor: a transmembrane glycoprotein containing an EGF domain and an immunoglobulin homology unit. Cell 69:559-572.

20 Wen, D., Suggs, S.V., Karunagaran, D., Liu, N., Cupples, R.L., Luo, Y., Janssen, A.M., Ben-Baruch, N., Trollinger, D.B., Jacobsen, V.L., Meng, S.-Y., Lu, H.S., Hu, S., Chang, D., Yang, W., Yanigahara, D., Koski, R.A., and 25 Yarden, Y. (1994) Mol. Cell. Biol. 14:1909-1919.

References to Example 6

1. Fischbach G.D. & Rosen KM ARIA: a neuromuscular 30 junction neuregulin. Annu Rev Neurosci. 20, 429- 458 (1997).
2. Marchionni M.A. et al. Neuregulins as potential neuroprotective agents. Ann. N. Y. Acad. ScL 825, 35 348-365 (1997)

3. Yang, X., Kuo, Y., Devay, P Yu. C . & Role, L.W. A cysteine-rich isoform of neuregulin controls the level of expression of neuronal nicotinic receptor channels during synaptogenesis. *Neuron* 20, 255-270 (1998).
4. Ozaki, M., Sasner, M., Yano, R . Lu, H.S., & Buonanno, A. Neuregulin-beta induces expression of an NMDA receptor subunit. *Nature* 390, 691-694 (1997).
5. Sandrock AW Jr. et al. Maintenance of acetylcholine receptor number by neuregulins at the neuromuscular junction in vivo. *Science* 276, 599- 603 (1997).
6. Marchionni M.A. et al. Gliai growth factors a real alternatively spliced erbB2 ligands expressed in the nervous system. *Nature* 362. 312-318 (1993).
7. Wen D. et al. Neu differentiation factor: a transmembrane glycoprotein containing an EGF domain and an immunoglobulin homology unit. *Cell* 69, 559-572 (1992).
8. Holmes W.E. et a/. Identification of heregulin. a specific activator of p185erbB2. *Science* 256, 1205-1210 (1992).
9. Falls, D.L, Rosen, K.M.. Corfas, G., Lane, W.S., & Fischbach, G.D. ARIA, a protein that stimulates acetylcholine receptor synthesis, is a member of the neu ligand family. *Cell* 72, 801-815 (1993).
10. Ho, W.H., Armanini M.P.. Nuijens. A., Phillips, H.S., & Osheroff, P.L. Sensory and motor

neuron-derived factor. A novel heregulin variant highly expressed in sensory and motor neurons. *J Biol Chem.* 270. 14523-14532 (1995).

5 11. Meyer D. et al. Isoform-specific expression and function of neuregulin. *Development* 124, 3575-3586 (1997).

10 12. Kramer R. et al. Neuregulins with an Ig-like domain are essential for mouse myocardial and neuronal development *Proc Natl Acad Sci USA* 93, 4833-4838 (1996).

15 13. Meyer D., & Birchmeier. C Multiple functions of neuregulin in development. *Nature* 378. 286-390 (1995).

20 14. Erickson S.L. et al ErbBS is required for normal cerebellar and cardiac development: a comparison with ErbB2-and heregulin-deficient mice. *Development* 124, 4999-5011 (1997).

25 15. Liu X. et al. Domain-specific gene disruption reveals critical regulation of neuregulin signaling by its cytoplasmic tail. *Proc. Natl. Acad. Sc./, in press.*

30 17. Hall Z.W. & Sanes J.R. Synaptic structure and development: the neuromuscular junction. *Cell* 72Suppl, 99- 121 (1993). 17. Naciff J.M., Misawa H., & Dedman J.R. Molecular characterization of the mouse vesicular acetylcholine transporter gene. *Neuroreport* 8, 3467- 3473 (1997).

35 18. Easter, S.S. Jr., Ross, L.S. & Frankfurter, A.

Initial tract formation in the mouse brain.
JNeurosci. 13, 285-299 (1993).

19. Jessen KR. & Mirsky R. Origin and early development
5 of Schwann cells. Microsc Res Tech. 41.
393-402 (1998)

20. Grinspan, J.B.. Marchionni. MA.. Reeves,
10 M., Coulaloglou, M., & Scherer, S.S. Axonal
interactions regulate Schwann cell apoptosis in
developing peripheral nerve: neuregulin receptors
and the role of neuregulins. J Neurosci. 16, 6107-
6118 (1996).

15 21. Riethmacher D. et al. Severe neuropathies in mice
with targeted mutations in the ErbB3 receptor.
Nature 389. 725-730 (1997).

22. Helliwell, T.R., Ellis, I.H., & Appleton, R.E.
20 Myotubular myopathy: morphological,
immunohistochemical and clinical variation.
Neuromuscul Disord. 8, 152-161 (1998).

23. Laporte J, et al. A gene mutated in X-linked
25 myotubular myopathy defines a new putative tyrosine
phosphatase family conserved in yeast. Nat Genet.
13: 175-182 (1996).

24. Trachtenberg J.T. & Thompson W.J. Schwann
30 cell apoptosis at developing neuromuscular junctions
is regulated by glial growth factor. Nature 379,
174- 177 (1996).

25. Oppenheim R.W. Cell death during development of the
35 nervous system. Annu Rev Neurosci. 14,

453-501 (1991).

26. Grieshammer, U., Lewandoski. M., Prevette, D., Oppenheim, R.W., & Martin, G.R. Muscle-specific cell ablation conditional upon Cre-mediated DNA recombination in transgenic mice leads to massive spinal and cranial motoneuron loss. *Dev. Biol.* 197, 234-247 (1998).

10 27. Jo. S.A., Zhu, X., Marchionni. M.A.. & Burden, S.J. Neuregulins are concentrated at nerve-muscle synapses and activate ACh-receptor gene expression. *Nature* 373. 158-161 (1995).

15 28. Moscoso LM. et al. Synapse-associated expression of an acetylcholine receptor-inducing protein. ARIA/hereregulin, and its putative receptors. ErbB2 and ErbB3, in developing mammalian muscle. *Dev Biol.* 172, 158-169 (1995).

20 29. DeChiara T.M. et al. The receptor tyrosine kinase MuSK is required for neuromuscular junction information in vivo. *Cell* 85, 501-512 (1996).

25 30. Gautam M. et al. Defective neuromuscular synaptogenesis in agrin-deficient mutant mice. *Cell*, 525-535 (1996).

30 31. Wang, F., Nemes, A., Mendelsohn. M., & Axel, R. Odorant receptors govern the formation of a precise topographic map. *Cell* 93, 47-60 (1998).

35 32. Streit A. et al. Preventing the loss of competence for neural induction: HGF/SF, L5 and Sox-2. *Development* 124, 1191-1202 (1997).

33. Karnovsky, M.J. & Roots L. A
"direct-coloring"thiocholine method for
cholinesterases. *J. Histochem. Cytochem.* 12, 219-221
5 (1964).

34. Lumsden, A. & Keynes R Segmental patterns of
neuronal development in the chick hindbram. *Nature*
337. 424-428 (1989).